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RESEARCH ARTICLE

Epidemiology of Bovine Hydatidosis: Urbanization, Dogs, Animal Care and Proximity to Slaughterhouses are Important Risk Factors for Cattle

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ABSTRACT

Cystic echinococcosis, a neglected tropical disease caused by Echinococcus granulosus, is of One-Health importance. The disease has significant impact on the economy of Pakistan, where livestock is an important pillar of farming. Given the large socio-economic and zoonotic importance of cattle, we explored echinococcosis prevalence in livestock, focusing on three previously little-studied districts of Punjab (Narowal, Sheikhupura and Sialkot), Pakistan. We screened in total 1168 slaughtered cattle for presence of hydatid cysts. The collected hydatid cysts were subjected to microscopy, histopathology, and PCR. Overall disease prevalence was 7.7% (n=1168), significantly higher in Narowal (9.6%) than in Sheikhupura (7.6%) and Sialkot (5.7%). The oldest cattle group (>5 years) had significantly higher prevalence (11.8%) than younger animals (6.8% in 3-5-year and 4% in 1-3-year-olds). Females had significantly higher prevalence (9.1%) than male (4.9%) cattle. Significantly more cysts occurred in cattle lungs (71.4%) rather than the liver (28.5%), and the number of fertile cysts was significantly higher in lungs (56.9%) compared to liver (50%). PCR and sequencing of one cyst confirmed the species to be Echinococcus granulosus, with phylogenetic analysis clustering our ND1 sequence with the G1-G3 lineages. We found hydatid cysts to cause histopathological changes in the host tissue surrounding the cysts, including atrophy, fibrosis, cell degeneration and leucocytic infiltration. General Linear Models revealed that animals (cattle and dogs) kept near slaughterhouses, particularly in urban and semi-urban areas, significantly increased the risk of cystic echinococcosis in cattle. Based on these findings, we recommend a public health campaign to increase awareness of zoonotic infections.

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INTRODUCTION

Livestock are affected by many infectious diseases that adversely reduce their growth and production. Cystic echinococcosis or hydatidosis is one such zoonotic disease, caused by the cestode parasite *Echinococcus* granulosus that infects both animals and humans (Ali *et* al., 2015). This is a neglected tropical disease (NTD) of One Health importance. The dog (*Canis familiaris*) is the definitive host for *Echinococcus granulosus*, whereas intermediate hosts are herbivores and omnivores including humans. The disease is transmitted to the intermediate host by ingestion of the cestode eggs expelled by the definitive host (Lawson and Gemmell, 1983). Humans act as accidental hosts by acquiring the infection through close contact with dogs, or by ingesting water or food contaminated with parasitic eggs (Nakao *et al.*, 2010; Torgerson *et al.*, 2011). In intermediate hosts, the parasite develops into a fluid filled hydatid cyst, typically in the liver and lungs and rarely in the kidney, brain and bone marrow, resulting in morbidity and mortality (Battelli 2009). In humans, the infection causes bile duct obstruction and pleural fistula disorders (Daali *et al.*, 2001).

echinococcosis Cystic has a cosmopolitan distribution (Haleem et al., 2018), and is highly prevalent in herd keeping areas across the world (Bekele and Butako 2011), ranging from 12% in India (Grakh et al., 2020), 13.9% in Iran (Vaisi-Raygani et al., 2021) and 22% in Ethiopia (Shumuye et al., 2021) to 53.9% in China (Fan et al., 2022; Yang et al., 2022). Within Pakistan, prevalence in different host species ranges from 2.4 to 65.4% (Tasawar et al., 2014). The parasite is responsible for huge economic losses due to reduced milk and meat production, and condemned meat (Lemma et al., 2014; Basinger et al., 2021). It causes an estimated loss of Rs 26.5 million annually to the livestock sector of Pakistan (Latif et al., 2010), approximately USD 1.65 per organ (Shafiq and Athar 2004). X-ray Computed Topography (CT) scans (e.g. Díaz-Menéndez et al., 2012), ELISA and PCR (Khan et al., 2023) can be exploited for diagnosis of this disease, but to reduce prevalence, further information is needed on the distribution and risk factors associated with E. granulosus.

There is limited data on the epidemiology of cystic echinococcosis in remote areas of Northern Punjab, Pakistan, as well as on the histopathological changes associated with this disease. The present study was thus designed to explore the prevalence, histopathology, and associated risk factors of the causative agent of bovine cystic echinococcosis while sampling slaughtered cattle in three districts of the Punjab Province in Pakistan.

MATERIALS AND METHODS

Ethical approval and sample size determination: All experimental procedures were approved by the Institutional Guidelines of Ethical Review Committee of UVAS, Lahore, vide letter No. 939-1, Dated 05-09-2019.

Samples size for determination of *Echinococcus* occurrence was calculated by considering previous prevalence of 45.4% (Shahzad *et al.*, 2014) with 95% confidence interval and 5% absolute precision. The formula used to calculate the sample size (Thrusfield 2018) was: $N = (1.96)^2 P (1-P)/d^2$, where N = required sample size, P is previous prevalence and d is desired absolute precision, such that N = (3.84) 0.45 (1-0.45) / (0.0025)=380. We collected 400 samples from each district to allow for any samples that might subsequently have to be excluded from data analysis.

Sample collection and questionnaire: From three districts in Northern Punjab, Pakistan (Narowal, and Sheikhupura Sialkot, Fig. 1), 1200 cattle (n=400/district)were examined in the main slaughterhouse for each district through random sampling for the presence of hydatid cysts in the liver, lungs, kidneys, and spleen between December 2019 to November 2020. The animals were categorised based on age as <1, 1-3, 3-5 and >5 years old. We subsequently excluded animals younger than 1 year from statistical analyses due to the small sample size (31), as cattle of this age are rarely slaughtered. Cysts were collected in sterilized containers with 70% ethanol (for DNA extraction) or 10% neutral buffered formalin (for histology). Samples for histopathology (individual cysts from eight different animals) were processed according to

Belina et al. (2015). A questionnaire based on 20 simple close-ended questions about owner and animal details was used for risk factor analysis. All 1168 farmers who completed the survey (one farmer for each surveyed cattle, and each animal/farmer in this study originated from a different farm) were asked about district of their farm, farm habitat (peri-urban, urban and rural), whether they kept dogs, reason for keeping dogs (guard dog, companion and hunting), if they dewormed their dogs, home slaughtering of animals, disposal of offal (buried, left open or undisclosed), feeding dogs with viscera, disposal of dog faeces, distance from abattoir, deworming of cattle. and animal feeding conditions (confined/mixed/grazing) in a face-to-face discussion at the slaughterhouses. We also recorded the species, age and sex of the animal, and season of the slaughter (spring - March, April, and May; summer - June, July, and August; autumn - September, October, and November; and winter - December, January, and February). Postslaughter, we recorded the presence of cysts (yes or no), cyst location (liver, lungs or other) and fertility (fertile, sterile, or calcified).

Parasite molecular identification: Hydatid cysts were characterized according to Haleem et al. (2018). To confirm parasite species, DNA was extracted from one randomly selected cyst sample using a commercially available DNA extraction kit (WizPrep[™] gDNA Tissue kit Wizbiosolutions, South Korea, W71060-300). A 226 bp fragment of the mtDNA ND1 gene was targeted by using primers Eg1F81 5'-GTT TTT GGC TGC CGC CAGAAC-3', Eg1R83 5'-AAT TAA TGG AAA TAA TAACAA ACT TAA TCA ACA AT-3' (Boufana et al., 2013). PCR was performed in a T100 Thermal Cycler (Bio-Rad, Hercules, CA, USA) as described previously (Mahmood et al., 2022). Briefly, a total reaction volume of 50 µL included 25 µL of 2X AmpMaster™ Taq master mix (GeneAll[®], Exgene[™], catalogue number 541-001), 10 µL Ultrapure[™] DEPC water (Invitrogen, 750023), 5 μ L each of the forward and reverse primer (50 μ M each), and 5 µL DNA extract, with PCR conditions: initial denaturation at 94°C for 3 minutes followed by 28 cycles (denaturation for 30 s at 94°C, annealing at 59.8°C for 30 s, extension at 72°C for 1 minute) and final extension step at 72°C for 5 minutes. The PCR product was then run on a 2% agarose gel (1 h at 120 V), stained with SYBR safe DNA (catalogue no. 2291850; Invitrogen, under Waltham, USA) and viewed MA, а transilluminator (Trans Lum SOLO, Biotop China, serial no. 21102053).

Sequencing and phylogenetics: The PCR product was sequenced by a commercial sequencing facility (1st BASE Pte Ltd., Singapore) using the forward primer. The resulting chromatogram was trimmed to delete low quality bases, and the resulting 162 bp sequence submitted to GenBank (Accession Number: OM935772). For comparison, we downloaded 13 ND1 sequences of *Echinococcus* from GenBank, covering the principal lineages of *Echinococcus granulosus* and closely related lineages (Bowles *et al.*, 1992). The downloaded ND1 sequences were aligned with our new sequence in Genious Prime (version 2022.2.2; www.geneious.com) using the

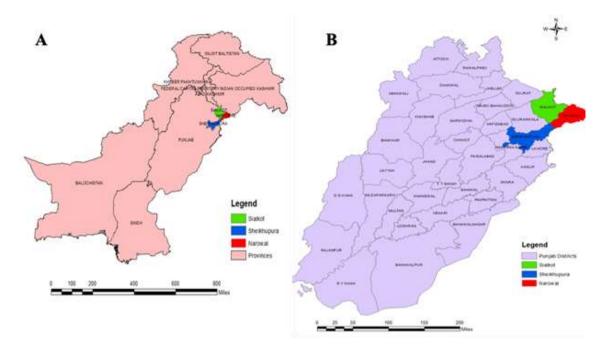


Fig. 1: Sampling districts in the current study. (A) Provinces of Pakistan and (B) districts within the province of Punjab. Maps constructed in QGIS software (3.28.1).

MUSCLE plugin. A maximum likelihood phylogenetic tree was reconstructed using IQ-tree webserver (version 2.2.0) (Trifinopoulos *et al.*, 2016). The most suitable substitution model for our alignment was determined by the built-in model finder function (yielding the HKY+I model with empirically determined base frequencies, based on the Bayesian Information Criterion) (Kalyaanamoorthy *et al.*, 2017). Statistical support for branches was determined from 1000 ultrafast bootstrap replicates (Hoang *et al.*, 2018).

Statistical analyses: All statistical analyses were performed using RStudio version 4.2.2. To understand the relationship between the chance of cysts being present within cattle and key environmental variables, we developed binomial generalized linear models (GLMs) with a logit link function, with the dependent variable being the presence or absence of cysts in slaughtered animals. We ran three models to analyse the data. In Model 1, the independent variables were district, age of animal and sex. Model 2 included habitat, deworming practice in cattle, farming type, animal feeding, home slaughtering, disposal of offal, seasons, and distance from abattoir as independent variables. Model 3 included keeping of dogs, reasons for keeping dogs, deworming of dogs, feeding dogs with viscera, and disposal of dog faeces. Removal of nonsignificant terms was performed to ensure model refinement (Thomas et al., 2017). The odds ratios were extracted from GLMs. Finally, Pearson Chi square tests assessed the association between cyst location and fertility. We excluded calcified cysts from the analysis, as only one calcified cyst was recovered.

RESULTS

Epidemiological study: From a total of 1168 slaughtered cattle from three districts in Pakistan, gross examination revealed 90 (7.7%) were infected with hydatid cysts (Fig.

2A and 2B). Infection rate was significantly higher in the Narowal district (9.6%) followed by Sheikhupura (7.6%) and Sialkot (5.7%) (Table 1). The oldest animals (>5 years) were significantly more prone to infection (11.8%) than those aged 3-5 years (6.8%) and 1-3-years (4%) (Table 1). Males were significantly less infected (4.9%) than females (9.1%) (P<0.001, Table 1). Cysts were more common in lungs (71.4%) compared to liver (28.5%), and cyst fertility was also significantly higher in lungs (56.9%) than liver (50%) (chi-square test, χ^2 =1203.7, P<0.05). Prevalence was significantly higher in winter (11.2%) compared to autumn (8.0%), spring (6.1%) and summer (5.4%) (Table 1).

The species of *Echinococcus* was confirmed through PCR and Sanger sequencing of one sample. Phylogenetic analysis (Fig. 3) revealed that the obtained ND1 sequence clustered with high statistical support (ultrafast bootstrap support: UF=100) within the wider G1/G3 lineage, identifying it as *E. granulosus sensu stricto* (Vuitton *et al.*, 2020). Our short alignment provided only limited resolution about clustering within this G1/G3 lineage, suggesting clustering of our sequence with moderate support (UF=74) with the G2/G3 types. The alignment however contained one diagnostic site that allowed us to distinguish between G1 and G3, and our sequence, G1 had a C, and all other sequences included in the alignment, including G3 and our sequence, had a T).

Sociodemographic survey regarding disease risk factors: Infection was significantly higher in urban (29.4%), and peri-urban (25.4%) areas compared to rural locations (5.4%; Table 1). Infection in cattle was significantly higher where farmers kept dogs at home or with other animals; (P<0.001) and higher when these were hunting (30.7%; P<0.27) or companion (69.3%; P<0.001) dogs compared to guard dogs (7.9%), and if the owners dewormed their dogs this decreased prevalence (P<0.001). The practice of home slaughtering did not increase the risk

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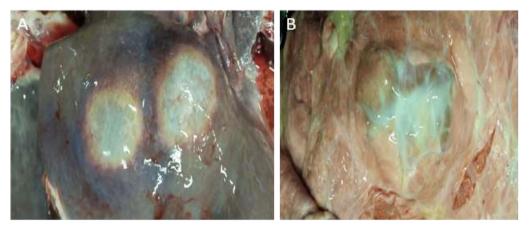


Fig. 2: Photographs showing multiple hydatid cysts present in the (A) liver and (B) lungs of cattle, encountered during gross examination at slaughterhouses.

Fig. 3: Maximum likelihood phylogeny of NDI sequences of main lineages of *Echinococcus granulosus* (*s.l.*) plus outgroups with GenBank accession numbers in brackets, along with the sequence obtained in the sequence obtained in the current study. Numbers on branches denote ultrafast bootstrap support values for the inferred groupings.

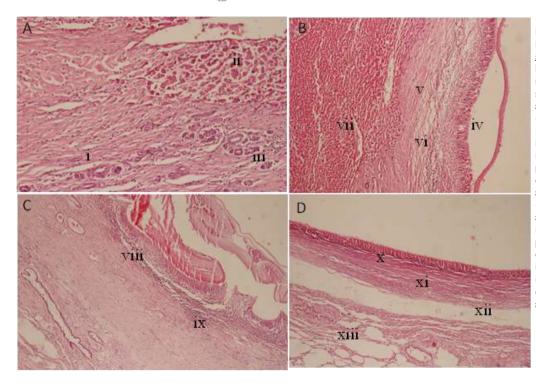


Fig. 4: Light microscope images of cattle tissue infected with Echinococcus granulosus stained with haematoxylin and eosin (40Xmagnification). (A) Liver showing (i) fibrosis, (ii) atrophy of hepatocytes and (iii) bile duct hyperplasia. (B) Liver with (iv) cyst lining, (v) infiltration of mononuclear inflammatory cells, (vi) proliferation of fibrous connective tissue and (vii) atrophy. (C) Lung showing (viii) degeneration of cells and (ix) leucocytic infiltration. (D) Lung with (x)proliferation of fibrous connective tissue, (xi) infiltration of mononuclear inflammatory cells, (xii) cystic wall and (xiii) atelectasis.

Table I: GLM analyses of prevalence of hydatidosis of slaughtered cattle and risk factors based on data from sociodemographic survey from farmers
completed in parallel to sample collection.

Risk Factors	Response	No. of Positive/ Total (%)	SE	Z. value	Odds ratios	P Value
Models I and 2: Risk factors	related to location and	d cattle. Reference categories Naro	wal and >5 yea	rs old, peri-u		10-20 km, buried
and autumn.		C	,	•		
	Narowal	38/395 (9.6)				
Districts	Sheikhupura	30/393 (7.6)	0.34	1.01	1.41	0.31
	Sialkot	22/380 (5.7)	0.40	0.55	1.24	0.58
	1-3	11/272 (4.04)	0.43	-3.70	0.20	0.001
Age (years)	3-5	38/541 (6.7)	0.35	-2.38	0.43	0.01
	>5	42/356 (11.79)				
Sex	Male	20/401 (4.97)	0.26	-3.23	0.42	0.001
	Female	70/767 (9.12)				
Habitat	Peri-urban	29/114 (25.43)				
	Rural	56/1037 (5.40)	0.40	-6.79	0.06	0.001
	Urban	5/17 (29.41)	0.67	-1.10	0.47	0.27
Feeding	Confined	23/192 (11.97)				
	Mixed	65/917 (7.08)	0.39	-3.15	0.28	0.001
	Grazing	2/59 (3.38)	0.98	-0.26	0.77	0.79
Deworming cattle	Yes	61/779 (7.83)	0.31	-1.55	0.61	0.12
Denorming cattle	No	29/389 (7.45)	0.51	1.55	0.01	0.12
Home slaughtering	Yes	49/409(11.98)	0.31	1.32	1.51	0.18
i tome staughter ling	No	41/759 (5.40)	0.51	1.52	1.51	0.10
Distance from abattoir	3-5 km	25/183 (13.66)	0.31	3.55	3.01	0.001
Distance if off abatton	5-10km	12/177 (6.77)	0.50	-2.43	0.29	0.01
	10-20km	53/808 (6.55)	0.50	-2.45	0.27	0.01
Disposal of offal	Buried	15/280 (5.35)				
Disposal of offai	Left open	34/129 (26.35)	0.33	5.66	6.50	0.001
	Unknown	41/759 (5.40)	0.31	0.02	1.0	0.97
Season of slaughter	Autumn	23/286 (8.04)	0.51	0.02	1.0	0.97
Season of slaughter		18/295 (6.10)	0.36	0.38	1.15	0.70
	Spring Summer		0.38	-0.14	0.94	0.88
	Winter	16/293 (5.46) 33/294 (11.22)	0.38	2.07	2.01	0.08
Madel 2: Diels franzens melanad						
		egories for reason of keeping dog v	was guard dog,	no dog tor o	eworming and	feeding viscera to
dogs and unknown for faeces of		(7/427/15 (0)	0.24	7.10	5.89	0.001
Keeping of dogs	Yes	67/427(15.69)	0.24	7.10	5.87	0.001
	No	23/741 (3.10)				
Reasons for keeping dogs	Guard dog	29/365 (7.94)	1.20	4.00		0.001
	Companion	34/49 (69.38)	1.29	4.99	6.34	0.001
	Hunting	4/13 (30.76)	2.36	1.09	1.88	0.27
	No dog	23/741 (3.10)	900.88	0.02	2.88	0.97
Deworming of dogs	Yes	41/363 (11.29)	1.33	3.85	5.79	0.001
	No	26/64 (40.62)				
	No dog	23/741 (3.10)				
Feeding dogs with viscera	Yes	52/80 (65)	1.15	4.10	1.12	0.001
	No	15/347 (4.32)				
	No dog	23/741 (3.10)				
Disposal of dog faeces	Yes	16/348 (4.59)	900.8	0.01	4.19	0.98
	No	51/79 (64.55)	900.8	0.02	1.84	0.98
	Unknown	23/741 (3.10)				

 Table 2: Prevalence of Echinococcus granulosus in cattle from two provinces (Punjab and KPK=Khyber Pakhtunkhwa) and different districts of Punjab, Pakistan.

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	Positive/total animals	Prevalence (%)	Reference
Province			
Punjab	90/1168	7.7	Current study
KPK	85/538	15.8	Haleem et al. 2018
КРК	41/189	21.7	Khan et al. 2021
District			
Narowal	38/395	9.6	Current study
Sheikhupura	30/393	7.6	Current study
Sialkot	22/380	5.7	Current study
Multan	105/1179	8.9	Mehmood et al. 2020
Sargodha	48/857	5.6	Mehmood et al. 2020
Islamabad & Rawalpindi	132/3845	3.4	Khan <i>et al.</i> 2020

of disease (P<0.18), whereas farmers who improperly disposed of offal (P<0.001) and/or fed dogs with viscera increased the risk of disease (P<0.001). The improper disposal of dog faeces did not increase the infection (P<0.98). Proximity of the home or dairy farm to an abattoir enhanced the risk of disease (P<0.001). There was a higher infection among confined cattle (23/192; 11.9%) compared to those which were kept in mixed conditions (65/917; 7.0%; P<0.001) or grazing (2/59; 3.3%; P<0.79). Surprisingly, deworming of cattle did not

significantly (P<0.12) impact prevalence of cystic echinococcosis.

Histopathology of liver and lungs: In the liver, fibrosis, atrophy of hepatocytes and bile duct hyperplasia were common, and at the cyst lining, infiltration of mononuclear inflammatory cells, proliferation of fibrous connective tissue and atrophy were seen (Fig. 4A and B). In the lungs, cellular degeneration, leucocytic infiltration, proliferation of fibrous connective tissue, infiltration of mononuclear

inflammatory cells, cystic wall and atelectasis were observed (Fig. 4C and D).

DISCUSSION

The livestock industry is threatened by many infectious diseases, including cystic echinococcosis that causes significant animal losses (Khan et al., 2023). In Pakistan, favourable socio-economic conditions for hydatidosis and high-level of infection in cattle mean that this is one of the most important diseases for cattle in the area (Fikire et al., 2012). We confirmed with NDI sequencing that *Echinococcus granulosus* is present in cattle in Northern Punjab, Pakistan. Phylogenetic analysis revealed clustering within the G1/G3 lineage (which includes the microvariant G2; Vuitton et al., 2020), with one substitution favouring a clustering with G3 rather than G1. Our findings indicate that the utilized primer pair, which was designed to be specific to the G1 lineage (Boufana et al., 2013), may also amplify the G2/G3 lineages, consistent with lineages G1, G2 and G3 being considered a single clade by Latif et al. (2010). We note, however, the limited phylogenetic resolution provided by our short fragment, so this finding should be reassessed with longer sequences. A recent phylogenetic study showed that although G1 and G3 are closely related, the two lineages are clearly diagnoseable with the resolution provided by whole mitogenome DNA sequences (Zhao et al., 2022). The G3 lineage has previously been reported in buffalo from India and China (Bowles et al. 1992; Guo et al. 2023), and the G1/G3 lineages in cattle from Pakistan (Mehmood et al., 2020), camels in Nigeria (Samari et al., 2022), and dogs, sheep, and humans in Uzbekistan (Kim et al., 2020). This highlights that the G1 and G3 lineages (at least those identified so far) are present in a variety of intermediate hosts and across a wide geographical distribution.

Within Pakistan, 0.71 million cattle suffer from echinococcosis in three North-East districts of the Punjab Province (Narowal, Sheikhupura and Sialkot), but at a lower prevalence (8%) than in North-West (Khyber Pakhtunkhwa; KPK) areas (Haleem et al., 2018; Khan et al., 2021; see Table 2). The Punjab is warmer than the KPK, and echinococcosis infection is negatively correlated with temperature; hence there is a lower risk of disease in warmer areas (Piarroux et al. 2015). The Punjab is also situated at a low altitude, and altitude is positively correlated with disease occurrence (Giraudoux et al., 2013). We speculate that the increased grassland area in the Punjab with lower cattle densities may result in the lower infection levels compared with the KPK. Other climatic variables and variable landscape features might also contribute to the difference in infection levels, but at a local level contact between animals and animal products is the most important risk factor (Hegglin and Deplazes, 2013).

The risk factors identified in this study indicate that keeping cattle close to a slaughterhouse enhances the risk of *E. granulosus* infection. Waste in the form of infected offal from the slaughterhouses can contaminate the surrounding environment for both final and intermediate hosts (Otero-Abad and Torgerson, 2013). Cattle from urban and peri-urban areas are also more likely to be infected with *E. granulosus* than those from rural habitats

(Acosta-Jamett et al., 2010). This may be a result of farmers in urban and peri-urban areas living in close contact with canines, the definite host for this parasite. Dogs in urban/peri-urban habitats have greater opportunity to ingest infected organs. Many butchers discard infected tissues (liver and lungs) inappropriately, increasing the risk for canids consuming this meat (Buishi et al., 2006). The rate of infection is higher in dogs whose owners feed them with viscera (Otero-Abad and Torgerson, 2013). Also, just keeping dogs with livestock increases the chance of echinococcosis (current study; Khan et al. 2020). Ingestion of eggs from contaminated soil is the primary route of echinococcosis infection for intermediate hosts (Shaikenov et al., 2003). In the current study, cattle maintained under mixed feeding or confined feeding conditions had a higher prevalence of disease, probably due to the higher risk of environmental contamination by dogs, compared to grazing cattle.

The liver and especially lungs were the most infected organs of cattle (current study; Khan et al., 2023). These highly vascularized organs are ideal for parasite growth, and the rich spongy nature of the lungs is probably more permissive for establishment and maintaining fertility of the oncosphere (Abunna et al., 2012). Not surprisingly, hydatid cysts cause histopathological changes to surrounding host tissues inducing atrophy, degeneration, and inflammatory cell infiltrations (current study; Beigh et al., 2017). Older animals (>5 years) had more cysts and higher prevalence (Khan et al. 2023), likely reflecting increased exposure to the parasite over time. In agreement to Mousa et al. (2015), the seasonal prevalence was higher in winter. The higher prevalence in female compared to male animals might be linked to pregnancy, parturition, and lactation, sometimes leading to malnutrition and transient immunosuppression thus enhancing their susceptibility to infection (Haleem et al., 2018).

Deworming of cattle did not appear to reduce the infection rate in the current study. This may be due to use of inappropriate drugs, incorrect administration, drug resistance (Gemmell et al., 2001), or cost. To reduce levels of echinococcosis, it is important to educate farmers about the timing and dosage of the correct cattle dewormers, and to ensure that they have access to effective, affordable dewormers. In agreement with (Mahmood et al., 2022), performing slaughtering at home did not increase the risk of infection in dogs, and reduced offal feeding to dogs reduced infection in cattle (current study; Wilson et al., 2019). Deworming of dogs can help control the disease in dogs (current study), which reduces environmental spread of Echinococcus eggs (Hegglin and Deplazes, 2013). Seminars/workshops should be arranged for dog owners and other members of the public to increase awareness of echinococcosis and other zoonotic infections.

Conclusions: Cystic echinococcosis, caused by *E.* granulosus, including lineage G1/G3, is prevalent in Northern Punjab, Pakistan (5.7-9.6%) and causes histopathological changes in vital organs. The disease shows significant association with host age and sex, district, and homes and dairy farms close to slaughterhouses. This study also highlights the disease risk of *E. granulosus* transmission between animals and humans if dogs are kept at home or with other animals. The zoonotic impact of

echinococcosis needs urgent attention by governments and stakeholders to reduce livestock loss and safeguard public health. A policy on dog keeping and handling, including registration advice on treatment, is needed, as well as control of stray dogs. Slaughterhouses with appropriate disposal pits and obligatory meat inspections are also highly recommended to reduce the prevalence of this common, but preventable, zoonotic disease.

Authors contribution: HA, MIR and MY designed the experiment. SK performed the field and laboratory experiments, analysed the data, and drafted the manuscript, with input from HA and JC. AI helped with sample collection. FH conducted phylogenetic analyses. HA, JC, and FH reviewed the manuscript.

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REFERENCES

- Ali I, Iqbal A, Munir I, et al., 2015. Molecular characterization of echinococcus species in Khyber pakhtunkhwa, pakistan. Acta Sci Vet 43:1-7.
- Abunna F, Fentaye S, Megersa B, et al., 2012. Prevalence of bovine hydatidosis in Kombolcha ELFORA abattoir, North Eastern Ethiopia. Sci Res 2:1-6.
- Acosta-Jamett G, Cleaveland S, Barend M, et al., 2010. Echinococcus granulosus infection in domestic dogs in urban and rural areas of the Coquimbo region, north-central Chile. Vet Parasitol 169:117-22.
- Basinger SC, Khan A, Ahmed H, et al., 2021. Estimation of the monetary burden of treated human cystic echinococcosis in Pakistan. Acta Trop 222:1-4.
- Battelli G, 2009. Echinococcosis: costs, losses and social consequences of a neglected zoonosis. Vet Res Commun 33:47-52.
- Beigh AB, Darzi MM, Bashir S, et al., 2017. Gross and histopathological alterations associated with cystic echinococcosis in small ruminants. | Parasit Dis 41:1028-33.
- Bekele J and Butako B, 2011. Occurrence and financial loss assessment of cystic echinococcosis (hydatidosis) in cattle slaughtered at Wolayita Sodo municipal abattoir, Southern Ethiopia. Trop Anim Health Prod 43:221-28.
- Belina D, Demissie T, Ashenafi H, et al., 2015. Comparative pathological study of liver fluke infection in ruminants. Indian J Vet Pathol 39:113-20.
- Boufana B, Umhang G, Qiu J, et al., 2013. Development of three PCR assays for the differentiation between Echinococcus shiquicus, E. granulosus (GI genotype), and E. multilocularis DNA in the coendemic region of Qinghai-Tibet plateau, China Am J Trop Med 88:795-802.
- Bowles J, Blair D and McManus DP, 1992. Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing. Mol Biochem Parasitol 54:165-73.
- Buishi I, Njoroge E, Zeyhle E, *et al.,* 2006. Canine echinococcosis in Turkana (north-western Kenya): a coproantigen survey in the

previous hydatid-control area and an analysis of risk factors. Ann Trop Med Parasitol 100:601-10.

- Daali M, ^Fakir Y, Hssaida R, *et al.*, 2001. Annales de chirurgie 2001. DOI: 10.1007/s00268-004-7516-z
- Díaz-Menéndez M, Pérez-Molina JA, Norman FF, et *al.*, 2012. Management and outcome of cardiac and endovascular cystic echinococcosis. PLOS Negl Trop Dis 6:1-8.
- Fan S, Dong H, Ma H, et *al.*, 2022. Meta-analysis on the prevalence of bovine hydatid disease in China from 2000 to 2021. Microb Pathog 168:1-11.
- Fikire Z, Tolosa T, Nigussie Z, et al., 2012. Prevalence and characterization of hydatidosis in animals slaughtered at Addis Ababa abattoir, Ethiopia. J Parasitol Vector Biol 4: 1-6.
- Gemmell M, Roberts M, Beard T, et *al.*, 2001. Control of echinococcosis. WHO/OIE manual on echinococcosis in humans and animals: a public health problem of global concern. Manual 286
- Giraudoux P, Raoul F, Pleydell D, et al., 2013. Drivers of Echinococcus multilocularis transmission in China: small mammal diversity, landscape or climate? PLOS Negl Trop Dis 7:1-12
- Grakh K, Prakash A, Mittal D, et al., 2020. Epidemiology, Risk Factors and Economics of Echinococcosis in India: A Review. Int J Livest Res 10:1-10.
- Guo B, Zhao L, Zhao L, et al., 2023. Survey and Molecular Characterization of Echinococcus granulosus sensu stricto from Livestock and Humans in the Altai Region of Xinjiang, China. Pathogens 12:1-134.
- Haleem S, Niaz S, Qureshi NA, et al., 2018. Incidence, risk factors, and epidemiology of cystic echinococcosis: a complex socioecological emerging infectious disease in Khyber Pakhtunkhwa, Province of Pakistan. Biomed Res Int 1-15
- Hegglin D and Deplazes P, 2013. Control of *Echinococcus multilocularis*: Strategies, feasibility and cost-benefit analyses. Int J Parasitol 43:327-37.
- Hoang DT, Chernomor O, Von Haeseler A, et al., 2018. UFBoot2: improving the ultrafast bootstrap approximation. Mol Biol Evol 35:518-22.
- Kalyaanamoorthy S, Minh BQ, Wong TK, et al., 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. Nat Methods 14:587-89.
- Khan A, Ahmed H, Simsek S, et al., 2020. Spread of cystic echinococcosis in Pakistan due to stray dogs and livestock slaughtering habits: research priorities and public health importance. Public Health Front 7:407-12.
- Khan J, Basharat N, Khan S, et al., 2021. Prevalence and molecular characterization of cystic echinococcosis in livestock population of the Malakand division, Khyber Pakhtunkhwa, Pakistan. Front Vet Sci 8:1-10.
- Khan S, Cable J, Younus M, et al., 2023. IEg67kDa Bovine Hydatid Cyst Antigen: a candidate for developing sero-diagnostic assays for cystic echinococcosis, a disease of One Health importance. Animals 13:866.
- Kim H-J, Yong T-S, Shin MH, et al., 2020. Phylogenetic characteristics of Echinococcus granulosus sensu lato in Uzbekistan. Korean J Parasitol 58:199-205.
- Latif AA, Tanveer A, Maqbool A, et al., 2010. Morphological and molecular characterisation of *Echinococcus granulosus* in livestock and humans in Punjab, Pakistan. Vet Parasitol 170:44-9.
- Lemma B, Abera T, Urga B, et al., 2014. Prevalence of bovine hydatidosis and its economic significance in Harar municipality abattoir, eastern Ethiopia. AEJSR 9:143-49.
- Lawson JR and Gemmell M, 1983. Hydatidosis and cysticercosis: the dynamics of transmission. Adv in Parasitol 22:261-308.
- Mahmood Q, Younus M, Sadiq S, et al., 2022. Prevalence and associated risk factors of cystic echinococcosis in food animals-a neglected and prevailing zoonosis. Pak Vet J 42:59-64
- Mehmood N, Arshad M, Ahmed H, et al., 2020. Comprehensive account on prevalence and characteristics of hydatid cysts in livestock from Pakistan. Korean J Parasitol 58:121-27.
- Mousa W, Mahdy O, Abdel-Wahab A, et al., 2015. Epidemiological and serological studies on cystic echinococcosis among camels in Egypt. J Parasitol Photon 105:212-18.
- Nakao M, Yanagida T, Okamoto M, et al., 2010. State-of-the-art Echinococcus and Taenia: phylogenetic taxonomy of humanpathogenic tapeworms and its application to molecular diagnosis. Infect Genet Evol 10:444-52.
- Otero-Abad B and Torgerson PR, 2013. A systematic review of the epidemiology of echinococcosis in domestic and wild animals. PLoS Negl Trop Dis 7:e2249.

Piarroux M, Gaudart J, Bresson-Hadni S, et al., 2015. Landscape and climatic characteristics associated with human alveolar echinococcosis in France, 1982 to 2007. Euro Surveill 20:21118.

- Samari H, Laurimäe T, Reghaissia N, et al., 2022. Molecular characterization of Echinococcus granulosus sensu lato genotypes in dromedary camels from extreme Sahara of Algeria based on analysis of nad2 and nad5 genetic markers. Act Trop 234:106616.
- Shafiq TA and Athar M, 2004. Epidemiology and economical aspects of hydatidosis in different animals, man and its control in sheep with indigenous plants; Ph.D. Thesis. University of the Punjab I-653.
- Shahzad W, Abbas A, Munir R, et al., 2014. A PCR analysis of prevalence of Echinococcus granulosus genotype G1 in small and large ruminants in three districts of Punjab, Pakistan. Pak J Zoo 46:1541-44.
- Shaikenov B, Torgerson P, Usenbayev A, et al., 2003. The changing epidemiology of echinococcosis in Kazakhstan due to transformation of farming practices. Act Trop 85:287-93.
- Shumuye NA, Ohiolei JA, Gebremedhin MB, et al., 2021. A systematic review and meta-analysis on prevalence and distribution of Taenia and Echinococcus infections in Ethiopia. Parasit Vectors 14:1-22.
- Tasawar Z, Naz F and Lashari M, 2014. The prevalence of hydatidosis in sheep and buffaloes at Multan, Punjab, Pakistan. Glob Vet 12:332-35.
- Thrusfield M, 2018. Veterinary Epidemiology, 4th edition; John Wiley & Sons: Edinburgh, UK. pp.887

- Thomas R, Vaughan I and Lello J, 2017. Data analysis with R statistical software: a guidebook for scientists. Newport: Eco Explore.
- Torgerson PR and Macpherson CN, 2011. The socioeconomic burden of parasitic zoonoses: global trends. Vet Parasitol 182:79-95.
- Trifinopoulos J, Nguyen L-T, Von Haeseler A, et al., 2016. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Res 44:232-35.
- Vaisi-Raygani A, Mohammadi M, Jalali R, *et al.*, 2021. Prevalence of cystic echinococcosis in slaughtered livestock in Iran: a systematic review and meta-analysis. BMC Infec Dis 21:1-10.
- Vuitton DA, McManus DP, Rogan MT, et al., 2020. International consensus on terminology to be used in the field of echinococcoses. Parasite 27:41.
- Wilson CS, Jenkins DJ, Brookes VJ, et al., 2019. An eight-year retrospective study of hydatid disease (*Echinococcus granulosus sensu stricto*) in beef cattle slaughtered at an Australian abattoir. Prev Vet Med 173:104806.
- Yang XB, Meng XZ, Zhao Y, et al., 2022. Meta-analysis of the prevalence of bovine cystic echinococcosis in China during decade. Res Vet Sci 152:465-75.
- Zhao Y, Gesang D, Wan L, et al., 2022. Echinococcus spp. and genotypes infecting humans in Tibet Autonomous Region of China: a molecular investigation with near-complete/complete mitochondrial sequences. Parasites & Vectors 15:75.