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RESEARCH ARTICLE

Study on the Acute and Sub-Acute Toxicity of Jia Wei San Huang Tang in Mice and Rats

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ABSTRACT

Jia Wei San Huang Tang (JWSHT) alleviates the cold properties of the original formula, San Huang Tang, and holds promise for treating gastrointestinal ailments. The primary objective of this study was to validate the safety of the JWSHT in a rat model and position the experimental foundation for future comprehensive investigations into its pharmacological effects and safe clinical application. Guided by the "Methodology of Pharmacological Research on Traditional Chinese Medicine," JWSHT underwent oral toxicity assessments. In the acute study, 60 Kunming mice (half male, half female) were categorized into five groups, receiving gavage doses of 16.0, 12.8, 10.2, and 8.2g/kg JWSHT for 7 days. Symptoms and mortality were recorded, and LD50 was calculated. To ascertain the maximum tolerated dose, mice received 128g/kg of the product and were observed for 14 days. In the subacute rat study, 80 rats were divided into three treatment groups and one control, administered doses of 16g/kg/d, 8g/kg/d, and 4g/kg/d for 30 days. Subsequently, rats were euthanized, and diverse parameters were analyzed to evaluate JWSHT's subacute toxicity. The acute toxicity test revealed that the LD50 was greater than 5g/kg. No signs of toxicity were observed in mice when administered at the maximum dose. The results of the subacute toxicity test indicated that the hemoglobin (HGB) levels in the high-dose group and the mean platelet volume (MPV) in the low-dose group were significantly higher than those in the control group (P<0.05). The alanine aminotransferase (ALT) levels in the low-dose group were significantly lower than the control group (P<0.05). The relative spleen weight in the male high-dose group was significantly higher than that in the control group (P<0.05). Mild bleeding was observed in the kidneys of the high-dose group, while other parameters showed no significant difference compared to the control group (P>0.05). Therefore, it was concluded that under the conditions of this study, the administration of JWSHT was relatively safe.

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INTRODUCTION

Chinese herbal medicine has a history of thousand years in the field of medicine, characterized by the synergistic effects of multiple components. By skillfully combining various herbal ingredients through precise proportions and administration methods, these formulations have demonstrated outstanding therapeutic efficacy (Lee *et al.*, 2022; Li *et al.*, 2022). Recorded in the Ming Dynasty's "Pocket Prescriptions," San Huang Tang (SHT) is a formula, primarily composed of *Coptis, Scutellaria baicalensis*, and *Phellodendron amurense*. SHT holds a significant position in the treatment of

gastrointestinal diseases, particularly in addressing dampheat diarrhea (Wang et al., 2014; Yin et al., 2021; Meng et al., 2022). Its historical application dates back centuries, and it continues to be widely used, providing effective treatment and relief to numerous patients. Although the effectiveness of classic herbal formulas is widely acknowledged, concerns about their safety arise when medical researchers continually modify these formulations, especially when introducing new herbal components, in pursuit of improved therapeutic outcomes and reduced side effects (Zhu et al., 2019; Cheng et al., 2022).

Through its mechanisms of anti-inflammatory, antibacterial, immunomodulatory and antioxidant actions,

SHT can alleviate various diseases such as diabetes, fatty liver, enteritis and cancer (Wu et al., 2019; Zhao et al., 2019; Chen et al., 2022; Wang et al., 2022). Additionally, it also effectively regulates the internal environment of the human body through exerting positive regulatory effects upon key physiological processes such as cellular metabolism and immune responses (Zhao et al., 2019; Tawulie et al., 2023). However, traditional SHT, due partly to its strong bitter taste and cold properties (Jin et al., 1995), may lead to potential side effects and discomfort with prolonged use, particularly affecting the digestive function, thus resulting in symptoms such as loss of appetite, abdominal distension, nausea, and vomiting. To optimize the efficacy of SHT and reduce its potential side effects, the original SHT formula was augmented with Chinese traditional medicinal herbs, Atractylodes Lancea, Fried Astragalus membranaceus, Prepared licorice, and Divine comedy to form a modified SHT as Jia Wei SHT (JWSHT). This purpose was to harmonize the properties of the SHT and potentially introduce new therapeutic effects. Literature shows that Astragalus membranaceus and Prepared licorice have been proven to enhance immunomodulation antioxidant effects (Chen et al., 2020; Leite et al., 2022), while the inclusion of Divine comedy and Atractylodes Lancea can balance the strong cold nature of SHT, thereby reducing its potential harm to the spleen and stomach (Qu et al., 2022; Liu et al., 2023). The goal of the formula of JWSHT is to construct a more comprehensive and effective treatment regimen that is also safer for patients. However, JWSHT is more complex in both its composition and effects, and this complexity might introduce new safety concerns. Therefore, to ensure the safe clinical application of JWSHT, this study utilized rat and mouse models to evaluate the toxicity levels of JWSHT, providing scientific evidence for its safety.

MATERIALS AND METHODS

Source of medicinal materials: Coptis (Dried rhizome of plants from the Ranunculaceae family, Sichuan, Batch No.: 22012503), Phellodendron amurense (Dried bark of the Phellodendron tree from the Rutaceae family, Anhui, Batch No.: 22040201), Scutellaria baicalensis (From the Lamiaceae family, Shanxi, Batch No.: 220105003), Atractylodes Lancea (Rhizome of the Atractylodes from the Asteraceae family, Anhui, Batch No.: 2112268), Fried Astragalus membranaceus (Roasted root of Astragalus from the Fabaceae family, Inner Mongolia, Batch No.: J210301), Prepared licorice (Processed dried root and rhizome of licorice from the Fabaceae family, Inner Mongolia, Batch No.: 211102), and Medicated leaven (A fermented mixture of Polygonum hydropiper, Artemisia annua, almond mud. adzuki beans, and fresh Atractylodes ear grass added to flour or bran, Sichuan, Batch No.: 202006082) were purchased from Tong Ren Tang pharmacy in Urumqi, The botanical materials China. morphologically identified in our laboratory and met the quality standards of the "Chinese Pharmacopoeia" (IPC, 2015).

Preparation and identification of JWSHT: According to the "Chinese Pharmacopoeia" (IPC, 2015), *Coptis*,

Phellodendron amurense, Scutellaria baicalensis, Atractylodes Lancea, Fried Astragalus membranaceus, Prepared licorice, and Divine comedy were ground into powder in a ratio of 10:10:10:15:15:6:10. The medicinal materials were soaked in distilled water at a 10 weight/volume (1:10, w/v) ratio for 30 minutes, then cooked for 2 hours and extracted twice. The filtrates were combined and concentrated to a raw medicine concentration of 1.6g/mL. For the experiments, the medicinal solution was diluted with distilled water to the desired concentration.

An appropriate amount of the powdered medicinal sample was weighed and added to 1 mL of 80% methanol, followed by ultrasonication for 10 minutes. It was then centrifuged at 14,000 rpm for 10 minutes. 0.8 mL of the supernatant was transferred to a centrifuge tube and centrifuged again. The resulting supernatant was placed into a sample vial. With a column temperature of 35°C and a flow rate of 0.2mL/min, the sample was separated on a chromatographic column. The separated compounds were ionized and introduced into a mass spectrometer. Subsequent mass spectrometric data collection was performed using the Q Exactive Plus Orbitrap (Thermo Fisher, Waltham, MA, USA) highresolution mass spectrometer. Both positive and negative ion modes were scanned simultaneously, with a scan range of m/z 100-1200. MS1 resolution was set to 70,000 and MS2 resolution was set to 17,500. The ion source voltage was 3.2kV, the capillary ion transfer tube temperature (Capillary temp) was 320°C, the auxiliary gas heating temperature (Aux gas heater temp) was 350°C, the sheath gas flow rate was 40L/min and the auxiliary gas flow rate was 15L/min. The AGC Target was set to 1e6, and the TopN was set to 5. The collision energy triggering MS2 scanning used a stepped fragmentation voltage NCE, set at 30, 40 and 50. Analysis was conducted using Compound Discoverer 3.3 software. Identification of each component was achieved by comparing the retention time, molecular weight (mass deviation <10 ppm) and MS2 fragment ions with the metabolites in the local mzVault database.

Animals and ethics: In this study, we utilized healthy Kunming (KM) mice, approximately 5 weeks old with a weight of around 20±2 g (both males and females, n = 100), and Sprague-Dawley (SD) rats, approximately 7 weeks old with a weight of around 200±20 g (both males and females, n = 80). The animals were sourced from the Animal Center of Xinjiang Medical University (2017013). They were housed in a room with a 12-hour artificial light cycle, at a temperature of 23±2°C, and a humidity of 50-65%. The animals were fed a standard diet and underwent an acclimatization period of 1 week. All animal experiments were conducted in accordance with ethical standards and were approved by the Ethics Committee of Xinjiang Agricultural University.

Acute oral toxicity study

Determination of median lethal dose (LD50): Based on the preliminary experimental results, the Hodge and Sterner method was employed. Sixty mice were randomly divided into five groups, with six mice in each group, evenly split between males and females. The mice were

fasted for 12 hours prior to dosing (without water deprivation) and were administered doses of 16.0, 12.8, 10.2, and 8.2g/kg via oral gavage. The control group received an equivalent volume of physiological saline. Observations were made continuously for 7 days, with checks conducted once in the morning and once in the afternoon, meticulously recording the mice's body weight, toxic reactions, and mortality. The LD50 was calculated using the modified Karber's method formula: LD50 = $1g^{-1}[Xm - i(\Sigma P - 0.5)]$, where Xm was the logarithmic value of the dose in the highest dose group; was the logarithmic value of the dose ratio; and ΣP was the sum of the mortality rates across all groups. If any mice died, a post-mortem examination was conducted. If no deaths occurred and further dosing was not feasible, a maximum dose test was performed.

Determination of maximum dosage: Forty mice were evenly divided into two groups, with 20 mice in each group, half male and half female. The mice were fasted for 12 hours but allowed access to water. Mice in the experimental group were administered the herbal compound at the Maximum Tolerated Dose Group (MTDG) (maximum permissible concentration of 1.6g/mL, 0.8mL per administration, dosed twice within 24 hours), while the control group received an equivalent volume of physiological saline. After oral administration, the mice were routinely housed for 14 days. Daily observations were made on the mice's mental state, and records were kept on their body weight, symptoms of poisoning, mortality rate, and time of death.

Sub-acute toxicity study: Based on the "Methodology of Pharmacological Research on Traditional Chinese Medicine"(Qi, 2006), the low dose in the subacute toxicity test was designed with reference to the clinical dose. The recommended dose of JWSHT for humans was 0.65g/kg. When converted to the rat dosage based on body surface area, it was approximately 4g/kg. Therefore, 4g/kg was chosen as the lowest administering dose for the subacute toxicity test. Subsequently, using the Hodge and Sterner method, SD rats, both female and male were divided into four groups, with 20 rats in each group (10 females and 10 males): High Dose Group (HDG) (16g/kg), Middle Dose Group (MDG) (8g/kg), Low Dose Group (LDG) (4g/kg), and Control Group (CG). Animals in each group were administered once daily at the allocated dose, with a gavage dose of 20mL/kg, continuously for 30 days. Daily records were kept on general behavior, clinical toxicity, mortality rate, and body weight. Cumulative weight gain (%) was calculated based on the initial weight. At the end of the dosing period, SD rats were fasted for 12 hours. Blood was drawn from the abdominal aorta. The rats were euthanized using an excessive amount of pentobarbital sodium, and various organs (heart, liver, spleen, lungs, stomach, duodenum) were excised and kidneys, weighed.

Hematological analysis: Blood was collected into anticoagulant tubes and analyzed using the ZC-980 Hematology Analyzer (Jilin Zichen Photoelectricity Technology Co., Ltd.).

Serum biochemistry analysis: Blood was collected into anticoagulant tubes and centrifuged at 1000 rpm for 3 minutes. Extract serum and place it into the Catalyst biochemical test kit (Adex Maine Bioproducts Trade Co., Ltd.). And used the PointcareM4 biochemical analyzer (Tianjin Mnchip Technology Co., Ltd.) to test four indicators: Creatinine (CREN), Blood urea nitrogen (BUN), Alanine Transaminase (ALT) and Aspartate Aminotransferase (AST).

Histopathological examinations: After weighing, the major organs (liver, heart, spleen, lungs, kidneys, stomach, and duodenum) from each group were immediately fixed in 4% formaldehyde (Gansu Weiboxin Biotechnology Co., Ltd.). After 24 hours, the tissues were dehydrated, embedded in paraffin, and sectioned into 4-5μm thick slices. The sections were then stained with Hematoxylin and Eosin (H&E) (Gansu Weiboxin Biotechnology Co., Ltd.) and observed under an IX53 inverted microscope (Olympus Corporation Co., Ltd, Japan) (Martey *et al.*, 2010; Afolabi *et al.*, 2012).

Statistical analysis: Data were presented as mean ± standard error of the mean (SEM). Statistical comparisons of the data were performed using the Statistical Package for the Social Sciences v26.0 for Windows (SPSS Inc., Chicago, IL, USA), including one-way or two-way analysis of variance, followed by a t-test to assess differences between groups. A P-value of less than 0.05 was considered statistically significant.

RESULTS

HPLC-MC analysis: The total ion chromatograms of JWSHT were generated using the Q Active Orbitrap high-resolution mass spectrometer in both positive and negative ion modes. Upon screening and analyzing the detected compounds, it was found that the modified San Huang Tang primarily contained 175 chemical components. These included flavonoids (60/175, 34%), alkaloids (25/175, 14%), acids (23/175, 13%), esters (11/175, 6%), terpenes (6/175, 3%), sugars (5/175, 3%), saponins (3/175, 2%), and some other types of compounds (Fig. 1, Table 1).

Acute oral toxicity

LD50 of JWSHT: After oral administration, two mice in the 12.8g/kg group exhibited symptoms of lethargy and disheveled fur immediately after gavage, but they recovered to their normal state after 6 hours. The rest of the mice appeared to be in good spirits and showed no adverse reactions. Continuous observation for 7 days revealed no abnormal reactions in any of the mice. There was no statistically significant difference in weight between the groups (P>0.05). Upon dissection, no abnormal pathological changes were observed in the major organs by the naked eye. The LD50 in this experiment was found to be greater than 5g/kg. According to toxicological evaluation standards and drug toxicity grading criteria (OECD, 2002), when the LD50 is greater than 5g/kg, the drug can be considered non-toxic.

Table 1: Chemical constituents of Jiawei San Huang Tang

Table 1: Chemical constituents of Jiawei San Huang Tang									
Name	Formula	Annot.	Calc. MW	m/z	RT		Reference Ion	Group Area:2	•
		DeltaMass	3		[min]	Best			Area:I
Berberine	C20 H17 N O4	[ppm]	335 1154	336.1227	24.566	Match 94.7	[M+H]+I	27048954291	17936393688
Dei Dei IIIe	C201117 IN O4	-1.02	333.1137	330.1227	24.300	70.7	fi.i.i.il.i	4.85	8.61
Palmatine	C21 H21 N O4	-0.43	351.1469	352.1542	24.396	90	[M+H]+I		52133013692
Coptisine chloride			319.0843	320.0916	23.361	89.9	[M+H]+I	51798087523	42738704700
Baicalin	C21 H18 O11			447.0921			[M+H]+I		33225869603
Epiberberine	C20 HI7 N O4			336.1227			[M+H]+I		28830114155
Wogonoside atrorrhizine	C22 H20 O11 C20 H19 N O4	-0.23		461.1077 338.1383		94.9	[M+H]+I [M+H]+I		28566871237 21539816085
(+)-Magnoflorine	C20 H23 N O4			342.1699		92.6	[M+H]+I	22462696670	
Phellodendrine chloride	02011231101	0.55		342.1699			[M+H]+I	20029661970	
Berberrubine	C19 H15 N O4	-0.35	321.1	322.1073	22.212	83.5	[M+H]+I	9885072298	8918333879
Wogonin	C16 H12 O5	-0.64	284.0683	285.0755	31.963	89.2	[M+H]+I	8164652843	11802509959
Baicalein	C15 H10 O5	-0.5		269.0455			[M-H]-I	7564244617	11336697169
2-Pyrrolidinecarboxylic acid L-	C5 H9 N O2	0.16		116.0706			[M+H]+I	7235441419	6204751550
Oroxylin A-7-O-β-D-glucuronide		-0.22 -0.65		461.1077 209.0807			[M+H]+I	6847516745	7351214728
4-Methylumbelliferone 7 Sucrose	C10 H8 O3 C12 H22 O11	-0.65 -0.49		341.1087		95.2	[M+H+MeOH]+I [M-H]-I	5734991496	5648860698
Genipin I-O-β-D-gentiobioside		-0.47	550.1897		18.746		[M+FA-H]-1	5372033696	304000070
Citric acid	C6 H8 O7	-0.27	192.027	191.0197		90.3	[M-H]-I	4637050242	2901982678
Chrysosplenetin B	C19 H18 O8	-0.59	374.I	375.1072			[M+H]+I	4038140086	5762896060
Liquiritin	C21 H22 O9	0.11		417.1192			[M-H]-I	3343108464	3295383608
Gardenoside	C17 H24 O11	-0.32	404.1317	449.1299	17.521	94.6	[M+FA-H]-I	3251411077	
Isoliquiritigenin	C15 H12 O4	-0.58		257.0807			[M+H]+I	2895003511	2649853485
Oxoglaucine	C20 HI7 N O5			352.1179			[M+H]+I	2791446081	2056453278
Chlorogenic acid	C16 H18 O9	-0.42		353.0877			[M-H]-I	2475604548	1815587361
Diammonium glycyrrhizinate	C42 H62 O16	-0.19 -0.2		823.4104	21.549		[M+H]+I ™ ⊔1 I	2198270174	1743600045
Liguiritigenin-7-O-β-D-apiosyl-4'-	C26 H30 O13	-0.2	330.1663	549.1613	21.547	03.1	[M-H]-I	2186304967	1754155883
O-β-D-glucoside Oroxylin A	C16 H12 O5	-0.69	284 0683	285.0755	32.761	86.9	[M+H]+I	2168268829	3123126262
Glycyrrhizic acid	C42 H62 O16	-0.01			31.22	93.3	[M-H]-I	1916467793	1278957722
18 β-Glycyrrhetintic Acid	C30 H46 O4	-0.55			31.221		[M+H-H2O]+1	1680769553	1254210380
Quinic acid	C7 H12 O6	-0.58	192.0633	191.056	20.673	90.4	[M-H]-I	1483898288	1205147638
Demethyleneberberine	C19 H17 N O4	-0.91	323.1155	324.1227	21.879	84.5	[M+H]+I	1219187093	1375367973
Shanzhiside		0.06	392.1319	391.1246	16.827	94.7	[M-H]-I	1123134865	
Trigonelline HCI	C7 H7 N O2	0.19	137.0477		1.558	89.3	[M+H]+I	1109410725	888224313.8
Taxifolin	C15 H12 O7	-0.1	304.0583		20.931		[M-H]-I	1049044418	1729196170
7-Methoxycoumarin Dihydropalmatine	C10 H8 O3 C21 H23 N O4	-0.66	353.1627	209.0807	18.746 22.581		[M+H+MeOH]+I		714134487.2
Manninotriose		0.07		549.1673		76.3 85	[M+H]+I [M+FA-H]-I	1020975116 946841215.6	679558877.1
Caffeic acid	C9 H8 O4	-0.36		163.0389			[M+H-H2O]+1	945005428	681077120.6
Geniposide	C17 H24 O10	-0.49	388.1368		19.605		[M-H]-I	943655172.4	10026172.01
L-Leucine L-	C6 H13 N O2	0.71	131.0947	132.102	4.101	71.3	[M+H]+I	938051582.2	1079188322
L-Glutamic acid L-	C5 H9 N O4	-0.14		295.1135	1.463	81.1	[2M+H]+I	886297461	808632738.2
Cryptochlorogenic acid	C16 H18 O9	-0.4		353.0877			[M-H]-I	885928381.6	360122304.8
Glabrolide	C30 H44 O4	-0.03	468.324	469.3312		76	[M+H]+I	827469382.6	665288845.8
Eriodictyol	C15 H12 O6 C22 H22 O10	-0.77 -0.54		271.0599 447.1283			[M+H-H2O]+I	746077057.1	777307737.4 467399260.1
Calycosin-7-O-β-D-glucoside Limonin	C26 H30 O8	-0.07	470.194	515.1922			[M+H]+I [M+FA-H]-I	664121966.3 662305995.9	767305049.5
5-Hydroxymethylfurfural 5-	C6 H6 O3	-0.07	126.0317			76.8	[M+H]+I	605782900.5	640474120.3
Formononetin	C16 H12 O4	-0.17		269.0808			[M+H]+I	598112212.8	538469631.8
Chrysin	C15 H10 O4	-0.24	254.0579	253.0506	32.203	91	[M-H]-I	581269945.4	756359267.5
Hydroxygenkwanin	C16 H12 O6	-1.06	300.0631	301.0703	25.036	71	[M+H]+I	575155484.1	779170624.9
Isoferulic acid	CI0 HI0 O4	0.23	194.058	195.0653			[M+H]+I	562985232.8	375178346
Dehydrocorydaline	C22 H23 N O4			366.1698			[M+H]+I	561603631.6	876104689.3
Shikimic acid	C7 H10 O5	-1.54		173.0453			[M-H]-I	524411878	377858590
Isoguanosine	C10 H13 N5 O5	-0.04	283.0917	282.0844	11./14	83.2	[M-H]-I	513943533.4	
Tetrahydropalmatine HCI	C21 H25 N O4	-0.68	355 1781	356.1854	22 17	73.9	[M+H]+I	497526284.6	227440095.8
Stachydrine	C7 HI3 N O2			144.1019		70	[M+H]+I	490994402.7	411619601.6
Scutellarin	C21 H18 O12		462.08	463.0872			[M+H]+I	477738701.8	453756349.9
Liquiritigenin	C15 H12 O4	-0.09		255.0663	25.195		[M-H]-I	470875444.2	498276154.3
Quillaic acid	C30 H46 O5	-0.16		469.3312		81.8	[M+H-H2O]+I	470663275.5	
Tetrahydropalmatine	C21 H25 N O4			356.1854			[M+H]+I	463273825.7	
Rutin	C27 H30 O16			609.1458			[M-H]-I	456659454.2	
p-Coumaric acid	C9 H8 O3	0.74		182.0813		83.6	[M+NH4]+I	455077115.9	470759340.1
Stachyose Morin	C24 H42 O21 C15 H10 O7	0.12 -0.02		665.2147 301.0354			[M-H]-I [M-H]-I	449038794.1 391442269.5	546752959.8 574739441.7
Crocetin	C20 H24 O4	-0.02		329.1747			[M+H]+I	389005369.7	J, 1/J/TT1./
Uridine	C9 HI2 N2 O6			243.0622		92.7	[M-H]-I	332445532.I	356751132.6
Puerarin	C21 H20 O9	0.25		415.1036			[M-H]-I	326208123	447255766.2
Calycosin	C16 H12 O5	-0.23		283.0612			[M-H]-I	314381652.7	460362314.4
Nicotinic acid	C6 H5 N O2	0.85		124.0394		82	[M+H]+I	284312516.2	362011207.2
Lawsone	C10 H6 O3	-0.1	1/4.0317	207.0652	23.707	/1.3	[M+H+MeOH]+I	281651795.8	

A 1:	60 1114 04	0.3	100 1040	107.0075	22.145	040	FN4 1 17 1	270005 475	2004101075
Azelaic acid	C9 H16 O4	-0.3		187.0975			[M-H]-1	278085475	300618197.5
Lysionotin	C18 H16 O7	-0.25		345.0968 123.0553			[M+H]+I	262076754.9	398114821.2
Nicotinamide Danshensu	C6 H6 N2 O C9 H10 O5	0.28 -0.52		123.0553		79.2	[M+H]+I [M-H]-I	246554136.1 240787784.4	391078122.4 680010015.7
Citropten	C11 H10 O4	-0.52		207.0652		79.8	[M+H]+I	237930222.2	000010013.7
Geniposidic acid	C16 H22 O10	-0.07	374.1213		17.06	94.1	[M-H]-I	221689420.5	
Maltopentaose	C30 H52 O26	-0.03		827.2674		88.7	[M-H]-I	197693522	165539139.9
Dehydrocostus lactone	CI5 HI8 O2	0.09	230.1307		33.477		[M+H]+I	191619291.5	220007612.3
Hispidulin	C16 H12 O6	-0.92		301.0703			[M+H]+I	172948812.6	442303372.2
Iristectorigenin B	CI7 HI4 O7	-0.13		331.0812			[M+H]+I	172587440.9	254596632.2
Atractylenolide II	C15 H20 O2	-0.29		233.1535		89	[M+H]+I	162189254.6	44729794.37
4-Methyl-6,7-dihydroxycoumari	C10 H8 O4	-0.51		225.0756	17.527		M+H+MeOH]+I		
7-Methoxy-4-methylcoumarin 7-	CII HI0 O3	-0.82	190.0628	191.0701	20.712	81.1	[M+H]+I	161683358.1	
Icaritin	C21 H20 O6	-0.05	368.126	367.1187	33.261	77.6	[M-H]-I	161297412.8	417463495
α-Linolenic acid α-	C18 H30 O2	-0.01	278.2246	279.2319	40.214	79.2	[M+H]+I	156275018.4	116332071.7
Salicylic acid	C7 H6 O3	-0.85	138.0316	137.0243	23.352	78	[M-H]-I	153535055.8	163314826.9
Cytosine	C4 H5 N3 O	0.02	111.0433	112.0505	2.698	84	[M+H]+I	147083023.7	153315462.5
Isoliquiritin	C21 H22 O9	0.24	418.1265	417.1192	24.098	88.7	[M-H]-I	133221996.5	138079214.6
Naringenin	C15 H12 O5	-0.64	272.0683	273.0756	22.996	88.3	[M+H]+I	130624336.2	117904472.3
Luteolin	C15 H10 O6	-0.34	286.0476	287.0549	23.923	80.3	[M+H]+I	109971069.5	174778031.7
Adenine	C5 H5 N5	-0.34	135.0545	134.0472	2.712	76.2	[M-H]-I	103489155.5	99665421.46
Orcinol gentiobioside	C19 H28 O12	0.09		447.1508			[M-H]-I	98328507.74	84109063.83
4-Methoxysalicylic acid 4-	C8 H8 O4	-0.95			17.063		[M-H]-I	96150418.34	64191364.66
2-Hydroxy-4-	C8 H8 O3	0.34	152.0474	153.0547	21.031	83	[M+H]+I	93414750.07	74368031.55
methoxybenzaldehyde 4-									
Naringenin chalcone	C15 H12 O5	-0.65		273.0756			[M+H]+I	92071848.33	52729117.94
Aurantio-obtusin	CI7 HI4 O7	-0.15		331.0812			[M+H]+1	91773643.82	126687175.2
Protocatechualdehyde	C7 H6 O3	-0.83		137.0243			[M-H]-I	89921778.39	83033068.2
3,5-Dicaffeoylquinic acid	C25 H24 O12	-0.05		515.1195		90.2	[M-H]-I	88697066.75	35552088.02
Coumarin	C9 H6 O2	0.08		147.0441	20.256		[M+H]+I	86443372.34	80413423.71
Isoscopoletin	C10 H8 O4	0.05		193.0496			[M+H]+I	84536549.74	F2002120.02
Astragaloside III	C41 H68 O14	0.47		829.4595			[M+FA-H]-I	79628037.82	52993130.82
Protocatechuic acid	C7 H6 O4	-0.59		153.0192			[M-H]-1	76692878.06	60771807.53
Sophoricoside	C21 H20 O10	0.26	432.1058		25.794		[M+H]+I	75004469.4	62533687.67
6"-O-Acetylglycitin 6"-O-	C24 H24 O11			489.1395			[M+H]+I	72476473.35	68973408.71
Dehydroglaucine	C21 H23 N O4 C23 H28 O10		353.1627	463.1612	23.057		[M+H]+I	71450813.55 69788361.94	70224355.61 70746902.5
Isomucronulatol 7-O-glucoside	C23 H24 O12			493.1342			[M-H]-I		99468240.84
Aurantio-obtusin β-D-glucoside							[M+H]+I	69407296.56	
L-Tryptophan L-	CII HI2 N2	-0.33	204.0070	203.0826	17.513	01.4	[M-H]-I	69369784.21	85898912.86
Wilforlide A	O2 C30 H46 O3	2.58	4E4 24E0	4EE 2E21	24 1 1 7	74 5	[M+LI]+I	(00225/5.40	E7007240 26
	C22 H30 O14	-0.23		455.3531 517.1561	34.117 19.869		[M+H]+I	68022565.49 65402334.62	57997240.36 77815112.77
Sibiricose A5 Astragalin	C21 H20 O11	0.23		447.0934			[M-H]- I [M-H]- I	62859237.67	21691640.5
Obacunone	C26 H30 O7	0.13		455.2065			[M+H]+I	60906559.58	88890122.13
Atractylenolide III	C15 H20 O3	-0.08		249.1485			[M+H]+I	56906183.47	64694686.67
3,4-Dihydroxyphenylethanol	C8 HI0 O3	-0.72			17.107		[M-H]-I	55363545.41	36704405.63
Isoalantolactone	C15 H20 O2	0.1		233.1536			[M+H]+I	55067911.68	82064542.7
Emodin-8-O-β-D-glucopyranoside		-0.19		431.0982			[M-H]-1		79498946.77
p-Hydroxybenzaldehyde	C7 H6 O2	-0.93		121.0294			[M-H]-I		64381018.63
Sinapic acid	CI I HI2 O5	1.41		225.0757			[M+H]+I	47498693.29	2115314.924
Oxyberberine	C20 HI7 N O5			352.1178			[M+H]+I	44266943.49	37221692.3
Retrochalcone	C16 H14 O4	-0.31		271.0964			[M+H]+I	43954859.8	36418976
Iridin	C24 H26 O13			521.1302		77.2	[М-Н]-1	41589249.35	64903825.96
Kaempferol	C15 H10 O6	-0.73		287.0548		73.3	[M+H]+I	40749305.51	32626254.75
Higenamine	C16 H17 N O3		271.1207		18.037		[M+H]+I	39851995.13	30705383.55
Verbascoside	C29 H36 O15	0.18	624.2055	623.1983	21.599	90.7	[М-H]-I	39803249.19	129856190.8
Licochalcone B	C16 H14 O5	-0.65	286.0839	285.0767	24.828	77.9	[M-H]-I	39680046.57	29486888.52
Dictamnine	C12 H9 N O2	0.12	199.0634	200.0706	29.588	70.I	[M+H]+I	37724673.2	30169953.83
Grosvenorine	C33 H40 O19	0.7	740.2169	739.2096	21.035	85.5	[M-H]-I	37555299.08	
Pinocembrin	C15 H12 O4	-0.07	256.0735	255.0663	32.482	85.4	[M-H]-I	34186278.5	43214161.16
Arglabin	C15 H18 O3	-0.13	246.1256	247.1328	29.249	74.2	[M+H]+I	33708994.45	37932066.06
Crocin II	C38 H54 O19	0.83	814.3266	859.3247	28.254	83.5	[M+FA-H]-I	30802562.89	
Loganic acid	C16 H24 O10	-0.27	376.1369	375.1296	17.597	70.5	[M-H]-I	30639261.02	6654104.213
Vicenin II		0.22		595.1657			[M+H]+I	30225338.05	25469962.06
Parthenolide	C15 H20 O3	-0.15		249.1485			[M+H]+I	29968185.43	29016137.84
5,7,3'-Trihydroxy-6,4',5'-	C18 H16 O8	-0.61	360.0843	361.0915	29.224	86	[M+H]+I	29264583.79	
trimethoxyflavone 5,7,3'-	OT 1				165		EN 4 1 15 ·	000=0=====	0.40=00
Gentisic acid	C7 H6 O4	-0.75		153.0192			[M-H]-I	29079702.89	24872015.23
Salidroside	C14 H20 O7	-0.07		345.1191			[M+FA-H]-I	28920199.62	16641012.27
Scutellarein	C15 H10 O6	-0.16		285.0404			[M-H]-I	27092104.56	57137458.67
Quercetin 3-O-β-D-Glucuronide		0.54	478.075	479.0824			[M+H]+I	27047028.43	16055904.91
Quercetin	CI5 HI0 O7	0.18		301.0354			[M-H]-1	26903655.53	2250207445
Benzoic acid	C7 H6 O2	0.31		123.0441			[M+H]+I	26165377.84	23592874.67
4-Hydroxybenzoic acid	C7 H6 O3	-0.8		137.0243		79.2	[M-H]-1	24315609.68	204710700
Alpinetin	C16 H14 O4	-0.31		271.0964			[M+H]+I	22254196.62	30471970.9
Artemetin	C20 H20 O8	-0.16	388.1158		35.298	74.2 73.6	[M+H]+I [M+⊔]+I	21879568.67	4923860.283
α-Boswellic acid α-	C30 H48 O3 C10 H8 O4	0.12 0		457.3678 193.0495			[M+H]+I [M+H]+I	21269491.59 20119662.39	5753882.64 40673439.02
Scopoletin	21011007	J	172.0723	173.0773	∠1.770	11.4	ferride.	2011/002.37	1007 373 7.02

Rutaevin	C26 H30 O9 -1.	4 486.1883	487.1956	28.615	76.7	[M+H]+I	19705260.6	23766931.62
Vicenin III	C26 H28 O14 -0.	.06 564.1479	563.1405	20.018	86	[M-H]-I	19208344.62	23504794.6
Pinosylvin	CI4 HI2 O2 -0.	.33 212.0837	213.091	25.595	91	[M+H]+I	18611822.09	23505540.68
5-Hydroxy-6,7-dimethoxylflavone	CI7 HI4 O5 0.4	15 298.0843	299.0916	27.118	70.3	[M+H]+I	15433097.65	22430875.6
Glycitin	C22 H22 O10 0.1	12 446.1214	445.1141	23.605	72.I	[M-H]-I	14216084.1	6224365.958
Curcumol	C15 H24 O2 -0.	.33 236.1776	237.1848	27.312	78.8	[M+H]+I	14031614.77	31922531.89
Rutaecarpine	C18 H13 N3 O -0.	28 287.1058	288.1131	34.492	76.7	[M+H]+I	12951521.75	9483861.964
5-O-Demethylnobiletin	C20 H20 O8 -0.	.08 388.1158	389.1231	29.359	74.9	[M+H]+I	12923763.3	29359440.16
Isoacteoside	C29 H36 O15 0.2	26 624.2056	623.1983	22.167	80.3	[M-H]-I	12280366.2	
Mulberrin	C25 H26 O6 0.0	9 422.173	423.1801	41.282	72.4	[M+H]+I	11741939.99	19936834.05
Glabridin	C20 H20 O4 -0.	.35 324.136	325.1433	36.463	71	[M+H]+I	2343825.143	4773490.293
Complanatuside	C28 H32 O16 0.1	2 624.1691	623.1621	22.537	72.3	[M-H]-I		19265628.05
Oroxin A	C21 H20 O10 -0.	.16 432.1056	431.0983	24.726	77	[M-H]-I		11800922.65
I-Caffeoylquinic acid I-	C16 H18 O9 -0.		353.0877	17.032	83.3	[M-H]-I		10605841.5
Irigenin	C18 H16 O8 0.0		359.0773	28.977	73.2	[M-H]-I		24080693.47
Oxysophocarpine	C15 H22 N2 0.1	16 262.1682	263.1755	17.05	77.I	[M+H]+I		20800299.93
	O2							
Pinoresinol 4-O-glucoside (+)-	C26 H32 O11 0.1		565.1926	22.309		[M+FA-H]-I		55356017.39
Betaine 甜菜碱	C5 HII N O2 0.0)2 117.079	118.0863	63.517	78.2	[M+H]+I		98924693.8
α-Cyperone α-	C15 H22 O 0.0	07 218.1671	219.1744	29.098	82.4	[M+H]+I		23793115.49
Eupafolin	C16 H12 O7 0.0	9 316.0583	317.0657	25.126	73	[M+H]+I		20430771.91
Tectorigenin	C16 H12 O6 -1.	28 300.063	301.0703	28.569	74.7	[M+H]+I		624186748.4
L-Phenylalanine L-	C9 HII N O2 0.3	39 165.079	166.0863	10.692	84.3	[M+H]+I		1201258069
Prim-O-glucosylcimifugin	C22 H28 O11 0.3	33 468.1633	469.1706	20.56	79.5	[M+H]+I		102820207.3
Isorhamnetin	C16 H12 O7 0.1	316.0583	315.0511	26.439	79	[M-H]-I		19222480.64
5,7-Dihydroxychromone 5,7-	C9 H6 O4 -0.	42 178.0265	177.0193	22.833	72.I	[M-H]-I		15981715.79
Dihydrosanguinarine	C20 HI5 N O4 -0.		334.1072			[M+H]+I		959789688.9
Chrysophanol 8-O-β-D-glucoside	C21 H20 O9 0.5	51 416.1109	415.1037	24.832	79.5	[M-H]-I		21387110.14
Emodin-3-methyl ether/Physcion	C16 H12 O5 -1.		285.0754	23.531	77.6	[M+H]+I		29119543.28
Baicalin methyl ester	C22 H20 O11 -0.		461.1077	24.894		[M+H]+I		87901531.48
Cimifugin	C16 H18 O6 -0.	84 306.1101	307.1176	22.011	86.5	[M+H]+I		165322440.8

Table 2: Blood routine test results of male rats in each group (n=10)

				(·· · · ·)
Items	control	High dose	Medium dose	Low dose
WBC(10 ⁹ /L)	6.22±1.41	6.64±1.12	7.04±1.08	6.62±1.14
RBC (10 ⁹ /L)	6.98±0.58	6.89±0.53	7.20±0.64	7.09±0.50
HGB (g/L)	144.7±5.03	151.80± 7.67*	145.2±5.07	144.4±4.50
HCT (%)	40.95±1.37	42.51±1.92	40.84±1.88	40.74±1.40
MCV (fL)	58.95±4.41	62.17±6.78	57.19±6.61	57.69±3.93
MCH (pg)	20.81±1.32	22.19±2.29	20.33±2.28	20.46±1.57
MCHC (g/L)	353.4±7.05	357.2±12.85	356.2±19.75	354.7± 12.01
RDW (%)	12.61±0.65	12.63±0.43	12.32±0.52	12.33±0.55
PLT (10 ⁹ /L)	922.4± 46.61	912.4±43.1	912.6±55.43	933.5± 40.95
PCT (%)	53.78±3.27	51.66±2.69	52.58±5.04	56.56±5.31
MPV (fL)	5.84±0.40	5.68±0.42	5.76±0.40	6.06±0.51*
PDW (%)	16.38±0.41	16.54±0.36	16.46±0.39	16.61±0.40
LYM (10 ⁹ /L)	4.98±0.81	5.37±0.98	5.50±0.72	5.17±1.05
MON(10 %L)	0.14±0.81	0.17±0.08	0.15±0.09	0.16±0.07
Gran (10 ⁹ /L)	1.45±0.27	1.43±0.30	1.56±0.31	1.56±0.31
LYM (%)	80.97±6.50	80.90±6.98	78.51±5.59	77.79±3.45
MON (%)	2.36±1.00	2.51±1.02	2.19±1.28	2.43±0.99
Gran (%)	23.66±3.35	21.73±4.23	22.16±2.57	23.54±1.93

The values are presented as means \pm standard errors of the mean (n = 10).

Determination of maximum dosage: The cumulative dosage administered to the mice within 1 day was 128g/kg. After the administration of the herbal medicine, the mice exhibited symptoms of lethargy due to stress. However, they resumed normal eating and activity within 4 hours. Over the course of 14 days, no deaths or pathological phenomena were observed in the mice. The variations in body weight of the experimental group were not statistically significant when compared to the control group (P>0.05) (Fig. 2). Upon dissection, no visible pathological changes were observed in the major organs. The maximum dosage of this herbal compound was determined to be 128g/kg.

Subacute oral toxicity

General observation and mortality: Daily oral administration of JWSHT at concentrations of 4, 8 and 16g/kg/day showed no significant behavioral changes in

the rats compared to the control group. The rats were alert, with even breathing patterns. On the second day after dosing, three rats from the middle dose group exhibited clustering behavior, ruffled fur, and lethargy. Additionally, one rat from this group showed symptoms of soft stool and other discomforts. However, by the third day, these symptoms had subsided and returned to normal. Throughout the experimental period, no rats died, resulting in a mortality rate of 0%.

Body weight and feed intake: After 30 days of oral administration in rats, there was no significant difference in body weight between the three herbal formula dose groups and the control group (P>0.05) (Fig. 2). There was no difference between males and females, indicating that the herbal formula had no significant impact on the growth and development of the rats.

Hematological parameters: Compared to the control group, the high-dose group showed an increase in hemoglobin (P<0.05), and the low-dose group exhibited a rise in mean platelet volume (P<0.05). However, there were no significant differences in other indicators between the various dose groups and the control group (P>0.05). (Table 2, 3).

Biochemical parameters: Compared to the control group, the alanine aminotransferase (ALT) levels in the low-dose group of rats decreased (P<0.05). The differences in the CREA, BUN, and AST indicators between each dosage group and the control group were not statistically significant. (P>0.05). (Table 4, 5).

Organ-to-body weight ratio: The results showed that after gavage, the relative spleen weight of the male high-dose group was significantly higher than that of the control group (P>0.05) (Fig. 3). There were no significant

^{*} Significance vs. the control group: P<0.05.

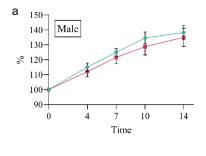
a 24% ■ Flavone (60/175) Alkaloid (25/175) 34% Acid (23/175) Ester (11/175) 2% Terpenes (6/175) 3% Saccharide (5/175) Saponin (3/175) Others (42/175) 14% 13% b

Fig. 1: The proportion of components(a) of JWSHT and TIC chromatograms under positive(b) and negative(c) ions modes.

differences in the relative weight of other organs (liver, heart, lungs, kidneys, stomach, and duodenum) compared to the control group.

Histopathologic analyses of vital organs: After gavage, the rats were dissected, and the size and color of the main organs were similar to those of the control group, with no

visible hemorrhagic spots, swelling, necrosis, or other gross pathological changes. Under the optical microscope, histopathological sections of rat tissues showed a small amount of red blood cell infiltration in the kidneys of individual rats in the high-dose group, but there were no significant differences in other organs compared to the control group. (Fig. 4).



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140

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90

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Male

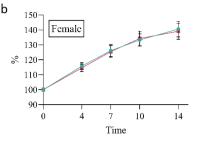


Fig. 2: Body weights of treatment groups that received acute (a, male; b, female) or subacute (c, male; D, female) treatment with JWSHT.

CG

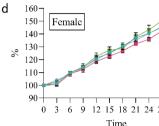
CG

HDG

MDG

LDG

MTDG



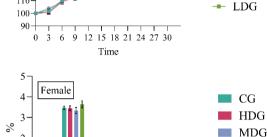


Fig. 3: Effect of Chinese herbal compound on organ-to-body weight ratio in rats(n=10), *Significance vs. the control group: P<0.05.

Male

Male

HRT LVR SPL LNG KID GI

12 15 18 21 24 27 30

Time

HRT LVR SPL LNG KID GI

Table 3: Blood routine test results of female rats in each group (n=10)

Items	control	High dose	Medium dose	Low dose
WBC (10 ⁹ /L)	6.64±1.34	6.87±1.17	6.29±1.40	6.60±0.91
RBC (10 ⁹ /L)	7.23±0.32	7.18±0.62	6.71±0.48	7.12±0.45
HGB (g/L)	144.2±4.98	150.00± 7.08*	144.5±4.28	145.7±3.43
HCT (%)	40.84±1.41	41.99±2.02	41.33±1.04	41.07±0.96
MCV (fL)	56.54±2.93	58.82±5.26	61.77±3.56	57.85±3.77
MCH (pg)	19.96±0.96	21.01±1.78	21.59±1.22	20.53±1.36
MCHC (g/L)	353.2± 10.52	357.5±15.29	349.6±5.24	354.9±9.08
RDW (%)	12.48±0.57	12.58±0.33	12.51±0.43	12.37±0.54
PLT (10 ⁹ /L)	917.7± 59.76	920.1±44.7	909.7±45.82	910.5± 39.78
PCT (%)	51.85±3.66	52.96±4.16	51.82±3.79	56.59±4.31
MPV (fL)	5.66±0.38	5.76±0.41	5.70±0.36	6.23±0.50*
PDW (%)	16.46±0.49	16.46±0.49	16.81±0.41	16.46±0.43
LYM (109/L)	5.26±1.25	5.51±0.85	5.70±0.36	5.25±0.98
MON(10 ⁹ /L)	0.17±0.08	018±0.08	0.16±0.07	0.16±0.05
Gran (109/L)	1.51±0.36	1.64±0.37	1.46±0.30	1.46±0.16
LYM (%)	78.96±5.79	80.63±7.01	80.31±6.99	79.16±5.04
MON (%)	2.60±1.24	2.59±1.05	2.65±1.19	2.52±0.99
Gran (%)	22.75±2.64	23.71±2.45	23.52±3.29	22.29±2.42

The values are presented as means \pm standard errors of the mean (n = 10). * Significance vs. the control group: P < 0.05.

Table 4: Blood biochemical test results of male rats in each group(n=10)

O. 2 - F (,			
Items	Control	High dose	Medium dose	Low dose
CREA	75.21±9.37	75.02±8.52	74.56±9.36	73.49±9.46
BUN	6.72±0.57	6.53±0.54	6.31±0.53	6.47±0.69
ALT	48.42±2.62	48.26±3.05	47.89±2.92	42.91±3.78*
AST	176.8±8.69	175.9±8.80	179.0±10.51	176.3±10.55

The values are presented as means \pm standard errors of the mean (n = 10). * Significance vs. the control group: P < 0.05.

Table 5: Blood biochemical test results of female rats in each group (n=10)

0 1	, ,			
Items	Control	High dose	Medium dose	Low dose
CREA	73.48±7.99	74.37±8.37	72.28±7.03	72.29±6.88
BUN	6.63±0.60	6.74±0.53	6.38±0.74	6.58±0.48
ALT	47.66±2.34	48.06±1.92	46.70±3.32	42.31±3.53*
AST	175.4±13.31	172.8±11.25	177.7±10.95	178.4±9.73
, <u>. </u>				

The values are presented as means \pm standard errors of the mean (n = 10). * Significance vs. the control group: P<0.05.

DISCUSSION

In recent years, medicinal plants have garnered significant attention due to their pharmacological effects. However, the toxicity of their active ingredients remains not fully elucidated. Their potential toxicity has become a serious medical concern. Therefore, this study aims to assess the acute and sub-acute toxicity of JWSHT, providing guidance for its safe clinical application.

The chemical complexity of JWSHT not only reveals its rich pharmacological effects but also provides crucial information for our assessment of acute and sub-acute toxicity. Through HPLC-MC technology, we identified that this formulation primarily contains 175 chemical components, especially flavonoids, alkaloids, and acids, which are well-known for their extensive pharmacological activities. The results show that flavonoids account for 34% of the total, alkaloids for 14%, and acids for 13%. Commonly found in plants, flavonoids have been proven to have antioxidant, anti-inflammatory, and anti-tumor effects (Imran et al., 2019; Singh et al., 2020; Chagas et al., 2022). Their antioxidant potential can mitigate cell damage caused by free radicals (Gupta et al., 2022; Carlini et al., 2022), while alkaloids exhibit analgesic and anti-inflammatory properties (Gao et al., 2022; Zhang et al., 2022), contributing to the overall safety of the drug. Therefore, the characteristics of these components are crucial for assessing the safety of JWSHT, and it is necessary to establish an HPLC-MC method to determine the structure and chemical properties of JWSHT.

In the acute study, we tested four dosage groups. The highest dosage (1.6g/kg) was the maximum concentration tolerated by mice. At the end of the experiment, no significant organ abnormalities were found during the

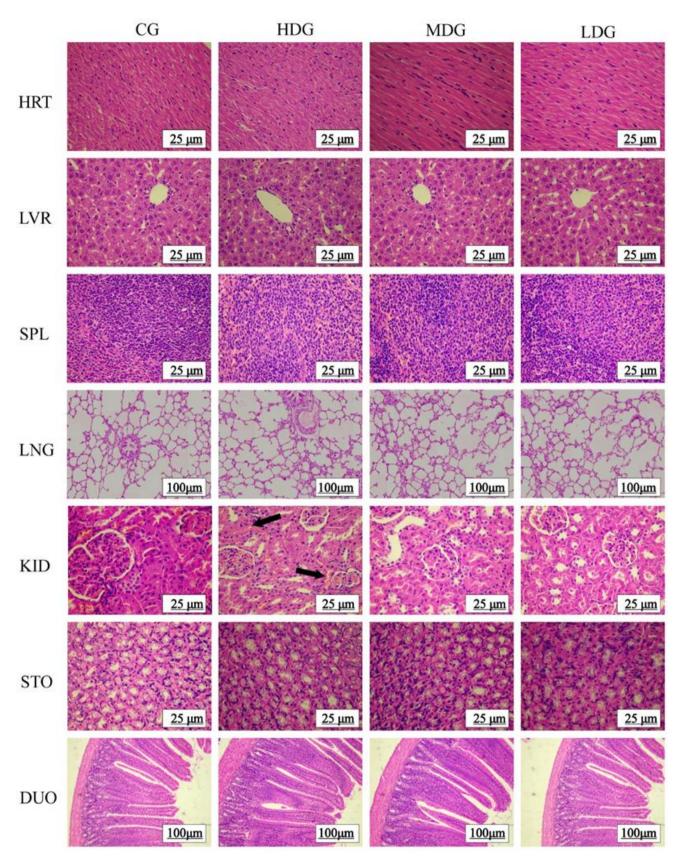


Fig. 4: Tissue sections of main organs of rats in each group. The arrows indicate a slight infiltration of red blood cells.

autopsy, and there were no obvious toxic reactions in mice. Therefore, it can be inferred that the LD50 of JWSHT in mice is much higher than 1.6g/kg. Further tests conducted at higher cumulative dosages also did not reveal any significant physiological or behavioral abnormalities. According to the OECD standards, with an

LD50 > 5g/kg, JWSHT can be classified as essentially non-toxic (OECD, 2002).

After completing the acute toxicity test, long-term toxicity tests are necessary. According to the dosage guidelines in the 'Methodology of Pharmacological Research on Traditional Chinese Medicine', the high-dose

group is designed based on clinical dosages to determine the safe dosage range for animals. Therefore, for this subacute toxicity test, 4g/kg was chosen as the low dosage group, and dosages were increased proportionally, divided into three dosage groups. After 30 days of continuous administration, there were no deaths in any of the treatment groups among the rats.

Body weight is considered a sensitive indicator of drug toxicity (Deyno et al., 2020; Wu et al., 2022; Canh et al., 2023), and no abnormal changes were observed in this study. Relative organ weight, especially the relative spleen weight, is considered an important indicator reflecting the immune function status of animals (Kang et al., 2021; Yan et al., 2021). The results showed that the relative spleen weight of the high-dose group was significantly higher than that of the control group, but no histopathological abnormalities were found. Astragalus polysaccharides, as the main component of Astragalus, can stimulate macrophage activity and increase the secretion of immune cell cytokines, thereby enhancing immune function (Li et al., 2022); Berberine, as the main component of Coptis, can affect the activation and secretion of lymphocytes, thereby regulating the immune system (Ehteshamfar et al., 2020). This suggests that JWSHT may cause an increase in spleen weight through immunomodulatory effects, indicative of enhanced immune response rather than pathological changes.

Additionally, from the hematological parameters, compared with the control group, the MPV value of the high-dose group significantly increased, suggesting enhanced activation and aggregation ability of platelets, and the HGB value of the high-dose group also showed an increasing trend, indicating an improvement in the quantity or quality of red blood cells. This could possibly be influenced by flavonoids and polysaccharides, as these substances can regulate platelet function by improving microcirculation or affecting components of the blood coagulation system (Zaragozá et al., 2021; Zaragozá et al., 2022; Araujo et al., 2023). These preliminary findings suggest that JWSHT may have an impact on the blood system, but the specific mechanisms and long-term effects require further study. Particularly, its impact on platelet function may need to be explored through more detailed experiments.

As a liver function marker, the decrease in ALT at a dosage of 4g/kg suggests that JWSHT might reduce liver metabolic capacity. However, no significant abnormalities such as hepatocellular degeneration or steatosis were observed in the histological examination of the liver (Xu et al., 2020; Choaib et al., 2023). Flavonoids and polyphenols have strong antioxidant effects, capable of mitigating cell damage caused by free radicals, thus protecting the liver (El-Aarag et al., 2019; Zhao et al., 2021). This suggests that JWSHT may have a potential protective effect on the liver, leading to reduced liver cell damage and consequently a decrease in ALT (Xu et al., 2018). However, given the observation of red blood cell infiltration in the kidneys of the high-dose group, we cannot entirely rule out the potential toxic risks of longterm administration or higher doses of JWSHT.

It is important to note that the components and mechanisms of action of traditional Chinese medicine compound formulations are very complex. There may be interactions between different components, and the ways in which they affect the body can vary greatly. This study provides preliminary evidence for the safety of JWSHT. Therefore, to ensure its safety in clinical applications, these preliminary results need to be validated and further explored through more extensive research.

Conclusions: The results of the acute test indicate that the LD50 of JWSHT is greater than 5g/kg, classifying it as essentially non-toxic. The sub-acute test results show no significant pathological changes after 30 days of administration at various doses. The study suggests that JWSHT is safe for clinical use.

Authors' contribution: WLY and CZ conceived the research idea, and both individuals made equal contributions. Professor WJQ developed the concept, monitored and mentored the proposal development. Professor YG. has polished this article. WLY \cdot CZ \cdot DZM and LZQ conducted the experiment. All authors have read and agreed to the published version of the manuscript.

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