



Relationships Between Thyroid Hormones, Serum Trace Elements and Erythrocyte Antioxidant Enzymes in Goats

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ABSTRACT

Thyroid hormones might be able to regulate the activities of superoxide dismutase (SOD), catalase and glutathione peroxidase (GPX) enzymes. The role of thyroid hormones in metabolic pathways and antioxidant enzyme activities are well known in many species. Nevertheless, there is no report describing probable relationship between thyroid hormones status, erythrocyte antioxidant enzymes and serum profiles of trace elements. This study was undertaken to investigate the relationship between these parameters in Iranian native goats. Blood samples were taken from the jugular vein of 50 clinically healthy Iranian native goats under aseptic conditions during 6 consecutive days of summer. The serum was analyzed for serum profile of thyroid hormones, trace elements, SOD and GPX activity. There were no significant differences in serum thyroid hormones, serum levels of zinc (Zn), copper (Cu), manganese (Mn), iron (Fe), selenium (Se) and antioxidant enzymes on different days ($P>0.05$). There were significant correlations between triiodothyronine (T_3) and GPX ($P<0.05$; $r=0.203$) and thyroxine (T_4) and GPX ($P<0.05$; $r=0.312$). There was no significant correlation between other parameters.

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INTRODUCTION

The mitochondrial antioxidant defense system is considerably influenced by the thyroid status of the body (Das and Chainy, 2001). Thyroid hormones seem to regulate the activities of superoxide dismutase (SOD), catalase and glutathione peroxidase (GPX) enzymes in the lymphoid organs and skeletal muscles (Pereira *et al.*, 1994).

The role of thyroid hormones in metabolic pathways and antioxidant enzyme activities are well known in many species such as rat (Asayama *et al.*, 1987) and camel (Zia-ur-Rahman *et al.*, 2007). Serum levels of thyroid hormones are mainly affected by general body metabolism (Yagil *et al.*, 1978), season (Nazifi *et al.*, 1999; Abdel-Magied *et al.*, 2000) and the water availability (Yagil *et al.*, 1978).

It is well known that steroid hormones, electrolytes and trace elements all play an important role in controlling the reproductive functions in animals (Al-Qarawi *et al.*, 2000). In camels, plasma testosterone concentrations have been found to be correlated significantly with the contents of Na^+ , K^+ , Ca^{++} and Mg^{++} in all genital organs but only

with epididymal contents of phosphorus and iron (Zia-ur-Rahman *et al.*, 2007). But the serum cholesterol level generally varies inversely with thyroid activity (Gueorguieva and Gueorguiev, 1997) and the concentrations of thyroid hormones do not correlate with cholesterol level in camels (Wasfi *et al.*, 1987), as in goat (Nazifi *et al.*, 2002). To the best of our knowledge, there is no report describing the probable relationship between thyroid hormone status, erythrocyte antioxidant enzymes and serum profiles of trace elements. Therefore, this study was undertaken to investigate the relationship between these parameters in Iranian native non pregnant goats.

MATERIALS AND METHODS

Experimental animals

The investigation was carried out on 50 non pregnant uniparous Iranian native goats which were reared mainly in South of Iran (Fars province). All the animals were clinically healthy and free from internal and external parasites. All the goats were treated with fenbendazol (10 mg/Kg) 30 days before the start of the study.

Blood sampling

Blood samples (10 ml) were taken from the jugular vein of experimental goats under aseptic conditions. The samples were taken at 8 a.m. during 6 consecutive days in summer, 2009 with a mean temperature of 38°C. For the determination of hemoglobin, superoxide dismutase (SOD) and glutathione peroxidase (GPX), blood samples were collected by jugular venepuncture into vacutainers containing ethylene diamine tetra acetic acid (EDTA) as an anticoagulant. For determination of serum thyroid hormones and trace elements, blood samples were collected into vacutainers, serum was separated by centrifugation at 750g for 15 min and stored at -20°C. The samples showing hemolysis were discarded.

Measurements

Serum triiodothyronine (T₃) and thyroxine (T₄) were measured by radioimmunoassay (RIA) method (kits available from Immunotech Company, IMMUNOTECH-Radiova-Prague-Czech Republic) in Namazi Research Center, Shiraz, Iran. The areas of validation for T₃ and T₄ assays included limits of detection and precision in standard curve following sample dilution, inter- and intra-assay coefficients of variation. The analytical sensitivity of T₃ and T₄ were 0.3 and <12.6 nmol/L, respectively. Intra- and inter-assays coefficients of variation for T₄ and T₃ were below 6.2 and 8.6%, and 3.3 and 8.6%, respectively.

Haemoglobin concentration was measured by cyanmethemoglobin method. SOD activity was measured by a modified method of iodophenyl nitrophenol phenyltetrazolium chloride (RANSOD kit, Randox Com, United Kingdom). This method employs xanthine and xanthine oxidase (XOD) to generate superoxide radicals which react with 2-(4-iodophenyl)-3-(4-nitrophenyl)-5-phenyltetrazolium chloride (INT) to form a red formazan dye. The superoxide dismutase activity was then measured by the degree of inhibition of this reaction. One unit of SOD was considered a 50% inhibition of reduction of INT under the condition of the assay. GPX was measured by the method of Paglia and Valentine (1967) (RANSEL kit, Randox Com, United Kingdom). GPX catalyzes the oxidation of glutathione (GSH) by cumene hydroperoxide. In the presence of glutathione reductase (GR) and NADPH, the oxidized glutathione (GSSG) is immediately converted to the reduced form with a concomitant oxidation of NADPH to NADP⁺. The decrease in absorbance was measured at 340 nm. For the measurement of serum trace elements, digestion of serum was performed by a mixture of perchloric and nitric acid (3:7 ratio), respectively. Manganese, copper, iron, selenium and zinc were measured using an atomic absorption spectrophotometer (Shimadzo AA-670, Kyoto, Japan).

Statistical analysis

The data were expressed in international units (SI) and analyzed by a repeated measure ANOVA and the Bonferroni multiple comparison test using SPSS/PC software (Norusis, 1993). Pearson's correlation coefficient was calculated for determination of the relationship between different biochemical markers during the consecutive days. All values were expressed as mean ±

standard error (SE) and P<0.05 was considered as statistically significant.

RESULTS

Diurnal variations of serum thyroid hormones (T₃ and T₄), antioxidant enzymes (SOD and GPX), and serum levels of trace elements (Zn, Cu, Fe, Mn and Se) in Iranian native goats during 6 consecutive days in summer are presented in Table 1. There were no significant differences in serum thyroid hormones, serum level of zinc (Zn), copper (Cu), manganese (Mn), iron (Fe), selenium and antioxidant enzymes among different days (P>0.05). There were significant correlations between T₃ and GPX (P<0.05; r=0.203) and T₄ and GPX (P<0.05; r=0.312). There was no significant correlation between other parameters.

DISCUSSION

In the present study, significant correlations between triiodothyronine (T₃) and GPX (P<0.05; r=0.203) and thyroxine (T₄) and GPX (P<0.05; r=0.312) were observed. There was no significant correlation between other parameters. The significant correlation between T₃ and GPX and T₄ and GPX are probably due to important role of thyroid hormones in lipid metabolism and antioxidant function of GPX in lipid peroxidation. The T₃ markedly affects lipid peroxidation and antioxidant enzyme activities in rat liver (Varghese *et al.*, 2001). It has been demonstrated with more accuracy that thyroid status controls the mitochondrial antioxidant defense system (Das and Chainy, 2001) by regulating the activities of SOD, catalase and GPX. Some studies have highlighted some complex relationships between thyroid status and antioxidant SOD and GPX activities. Asayama *et al.* (1987) suggested that increased lipid peroxidation in hyperthyroid rats was linked to enhanced oxidative metabolism and decreased GPX activity, whereas Mano *et al.* (1995) observed increased SOD and GPX activities in hyperthyroid rats compared to euthyroid animals. It was stated that GPX activity was increased, while glutathione concentrations remained unaltered in both hyperthyroid and hypothyroid rats (Shinohara *et al.*, 2000; Sawant *et al.*, 2003). Thyroid hormones may increase the activity of SOD and decrease GPX (Pereira *et al.*, 1995). It seems as if the T₄:T₃ ratio is more important than the level of individual hormone (Zia-ur-Rahman *et al.*, 2007), and it might be influenced by the season, temperature and effect of seasonal variation in the feed supply (Fay *et al.*, 2003).

Normal thyroid status is dependent on the presence of many trace elements for both the synthesis and metabolism of thyroid hormones. Selenium is required for conversion of thyroxine (T₄) into the more active triiodothyronine (T₃) via the enzyme type 4 deiodinase (Awadeh *et al.*, 1998). Additionally, selenoperoxidases and thioredoxin reductase protect the thyroid gland from peroxides produced during the synthesis of hormones (Aurthor and Beckett, 1999). However, there are some other trace elements such as iron, zinc and copper that their role in the thyroid is less well defined but sub-optimal dietary intakes of these elements can adversely affect thyroid hormone metabolism (Aurthor

Table 1: Variation in the concentrations of serum thyroid hormones, trace elements and antioxidant enzymes in Iranian native goats during 6 days (n = 50)

Days	Parameters								
	T ₃ (nmol/L)	T ₄ (nmol/L)	Zinc (µmol/L)	Copper (µmol/L)	Iron (µmol/L)	Manganese (µmol/L)	Selenium (µmol/L)	SOD (U/g Hb)	GPX (U/g Hb)
1 st	1.40 ± 0.13	70.35 ± 2.38	21.48 ± 2.55	19.08 ± 0.67	14.61 ± 1.07	0.03 ± 0.004	0.15 ± 0.02	1148.20 ± 77.79	303.07 ± 3.94
2 nd	1.36 ± 0.11	67.27 ± 2.72	19.76 ± 2.93	20.61 ± 0.93	15.62 ± 1.17	0.03 ± 0.005	0.16 ± 0.03	1167.43 ± 81.24	314.71 ± 5.73
3 rd	1.42 ± 0.14	73.29 ± 2.65	20.56 ± 2.38	20.74 ± 0.89	13.97 ± 1.36	0.04 ± 0.006	0.18 ± 0.04	1192.33 ± 71.37	316.27 ± 7.30
4 th	1.35 ± 0.13	71.94 ± 2.11	23.49 ± 2.87	19.83 ± 1.09	14.45 ± 1.02	0.04 ± 0.007	0.15 ± 0.03	1210.17 ± 85.26	317.83 ± 6.62
5 th	1.43 ± 0.17	65.34 ± 2.88	22.32 ± 2.63	18.96 ± 1.07	14.21 ± 1.19	0.03 ± 0.006	0.17 ± 0.04	1207.72 ± 78.87	311.65 ± 6.93
6 th	1.41 ± 0.15	69.47 ± 2.56	21.87 ± 2.89	18.87 ± 1.13	14.73 ± 1.12	0.04 ± 0.006	0.14 ± 0.05	1138.39 ± 83.93	319.53 ± 7.21

Values are presented as mean ± SEM. There were no significant differences in serum thyroid hormones, trace elements and antioxidant enzymes in different days (P>0.05).

and Beckett, 1999). Nevertheless, we couldn't find any significant correlation between trace elements, thyroid hormones and antioxidant enzymes. Interrelationships among copper and iodine and thyroid hormones were studied in rats by Aurthor *et al.* (1996). Kecici and Keskin (2002) reported a significant negative correlation between zinc concentrations of erythrocytes and serum thyroid hormones in healthy male Herino lambs and Angora goats. Copper deficient rats showed a decrease in the value of iodine metabolism in different organs and tissues excluding liver, whereas a sharp increase in the content of organic iodine was observed. In fact, copper deficiency enhances the effect of hypothyroidism (Aurthor *et al.*, 1996). Wichtel *et al.* (1996) showed that the plasma concentration of total thyroxin was increased (P<0.001) by selenium treatment and Bik (2003) determined the effect of selenium and iodine oral supplements on the concentrations of T₃ and T₄ in the serum of sheep. It is important to note that only when selenium levels decreased by more than 80%, deiodinase activity was markedly decreased (Bates *et al.*, 2000). Bates *et al.* (2000) stated that with the exception of liver, skin and non pregnant uterus, all of the tissues studied (including cerebrum, thyroid, pituitary, brown adipose tissue, ovary, testes and placenta) maintained substantial deiodinase activity (>50%) during prolonged selenium deficiency. Although the ability of a tissue to maintain deiodinase activity in the face of dietary selenium deprivation was associated in some tissues with a concomitant local preservation of selenium concentration, this was not the case for all tissues. How selenium levels are maintained in specific tissues, whether selenium is sequestered in specific cells of a tissue or organ during dietary selenium deprivation and the precise mechanism by which plasma T₃ levels are maintained in selenium deficient animals remain unanswered (Bates *et al.*, 2000).

Copper is the main component of SOD that plays a vital role as an antioxidant and protects the testis from oxidative stress (Henkel *et al.*, 2003). Humphries *et al.* (1983) revealed that in experimental copper deficiency in calf, plasma concentration of copper and SOD activity of erythrocytes severely decreased. Similarly, Konstantinova

and Russanov (1988) found a positive correlation between plasma concentration of copper and SOD activity of erythrocytes. However, we could not show any correlation between copper concentration and SOD activity in Iranian native goats. Inversely, there was a negative correlation between these parameters in human as previously stated (Tungtrongchitr *et al.*, 2003). We could not find any correlation between zinc concentration and SOD activity either, but it is known that zinc is an essential element that controls the balance in oxidant-antioxidant system (Brighthope, 2004).

It is not possible to explain fully results of the present study at this moment of time. Further investigations are needed to interpret these changes.

REFERENCES

- Abdel-Magied EM, AA Taha and AB Abdalla, 2000. Light and electron microscopic study of the thyroid gland of the camel (*Camelus dromedarius*). Anat Histol Embryol, 29: 331–336.
- Al-Qarawi AA, HA Abdel-Rahman, MS El-Belely and SA El-Mougy, 2000. Age-related changes in plasma testosterone concentrations and genital organs content of bulk and trace elements in the male dromedary camel. Anim Reprod Sci, 62: 297–307.
- Asayama K, K Dobashi, H Hayashibe, Y Megata and K Kato, 1987. Lipid peroxidation and free radical scavengers in thyroid dysfunction in the rat: a possible mechanism of injury to heart and skeletal muscles in hyperthyroidism. Endocrinology, 121: 2112–2118.
- Aurthor JR and GJ Beckett, 1999. Thyroid function. British Med Bull, 55: 658–668.
- Aurthor KA, M Kirchgessner and K Eder, 1996. Concentrations of thyroid hormones in serum and activity of hepatic 5 monodeiodinase in copper-deficient rats. Zeitschrift Fur Ernährungswissenschaft, 35: 288-291.
- Awadeh FT, RL Kincaid and KA Johnson, 1998. Effect of level and source of dietary selenium on

- concentrations of thyroid hormones and immunoglobulins in beef cows and calves. *J Anim Sci*, 76: 1204-1215.
- Bates JM, VL Spate, JS Morris, DL Stgermain and VA Calton, 2000. Effect of selenium deficiency on tissue selenium content, deiodinase activity and thyroid hormone economy in the rat during development. *Endocrine*, 141: 2490-2500.
- Bik DE, 2003. Influence of selenium and iodine supplementation on thyroid hormone concentrations in blood serum of sheep. *Bull Vet Inst Pula*, 59: 1126-1129.
- Brighthope I., 2004. The therapeutic potential of antioxidants in prevention and treatment of degenerative diseases. *Aust J Nutr Environ Med*, 13: 15-25.
- Das K and GB Chainy, 2001. Modulation of rat liver mitochondrial antioxidant defense system by thyroid hormone. *Biochem Biophys Acta*, 1537: 1-13.
- Fay B, M Bengoumi, F Moutaouakil and F Farge, 2003. Seasonal variations of the plasma thyroid hormone concentrations and the body temperature in the dromedary camel. *J Camel Prac Res*, 10:115-119.
- Gueorguieva TM and IP Gueorguiev, 1997. Serum cholesterol concentrations around parturition and in early lactation in dairy cows. *Révue Méd Vét*, 148: 241-244.
- Henkel R, C Baldauf and WB Schill, 2003. Resorption of the element zinc from spermatozoa by the epididymal epithelium. *Reprod. Dom Anim*, 38: 91-101.
- Humphries WR, M Philippo, BW Young and I Bremner, 1983. The influence of dietary iron and molybdenum on copper metabolism in calves. *British J Nutr*, 49: 77-86.
- Kececi T and E Keskin, 2002. Zinc supplementation decreases total thyroid hormone concentrations in small ruminants. *Acta Vet Hung*, 50: 93-100.
- Konstantinova SG and EM Russanov, 1988. Effect of pregnancy and fetal development on sheep liver superoxide dismutase activity. *Res Vet Sci*, 45: 287-290.
- Mano T, R Sinohara, Y Sawai, N Oda, Y Nishida, T Mokumo, K Asano, Y Ito, M Kotake and M Hamada, 1995. Changes in lipid peroxidation and free radical scavengers in the brain of hyper and hypothyroid aged rats. *J Endocrinol*, 147: 361-365.
- Nazifi S, HR Gheisari and H Poorabbas, 1999. The influence of thermal stress on serum biochemical parameters of dromedary camels and their correlation with thyroid activity. *Comp Haematol Inter*, 9: 49-54.
- Nazifi S, HR Gheisari and F Shaker, 2002. Serum lipids and lipoproteins and their correlations with thyroid hormones in clinically healthy goats. *Vet Arhiv*, 72: 249-257.
- Norusis MY, 1993. SPSS for Windows Base System. Users Guide Release 6.0. 1st Ed, SPSS Inc. Michigan, USA, PP: 281-290.
- Paglia DE and WN Valentine, 1967. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *J Lab Clin Med*, 70: 158-169.
- Pereira B, LF Rosa, DA Safi, EJ Bechara and R Curi, 1994. Control of superoxide dismutase, catalase and glutathione peroxidase activities in rat lymphoid organs by thyroid hormones. *J Endocrinol*, 140: 73-77.
- Pereira B, LF Rosa, DA Safi, EJ Bechara and R Curi, 1995. Hormonal regulation of superoxide dismutase, catalase and glutathione peroxidase activities in rat macrophages. *Biochem Pharmacol*, 50: 2093-2098.
- Sawant BU, GD Nadkarni, UR Thakare, LJ Joseph and MG Rajan, 2003. Changes in lipid peroxidation and free radical scavengers in kidney of hypothyroid and hyperthyroid rats. *Indian J Exp Biol*, 41: 1334-1337.
- Shinohara R, T Mano, A Nagasaka, R Hayashi, K Uchimura, I Nakano, F Watanabe, T Tsugawa, M Makino, H Kakizawa, M Nagata, K Iwase, Y Ishizuki and M Itoh, 2000. Lipid peroxidation levels in rat cardiac muscles are affected by age and thyroid status. *J Endocrinol*, 164: 97-102.
- Tungtrongchitr R, P Pongpaew, B Phonrat, A Tungtrongchitr, D Viroonudomphol, N Vudhivai and FP Schelp, 2003. Serum copper, zinc, ceruloplasmin and superoxide dismutase in Thai overweight and obese. *J Med Assoc Thailand*, 86: 543-551.
- Varghese S, PS Lakshmy and OV Oommen, 2001. Changes in lipid peroxidation and antioxidant enzyme activities by triiodothyronine (T₃) and polyunsaturated fatty acids (PUFA) in rat liver. *Endocrinol Res*, 27: 409-416.
- Wasfi IA, AM Hafez, FMA El-Tayeb and AY EL-Taher, 1987. Thyroid hormones, cholesterol and triglyceride levels in the camel. *Res Vet Sci*, 42: 418.
- Wichtel JJ, KG Thompson, AL Craigle and NB Williamson, 1996. Effects of selenium and iodine supplementation on the growth rate, mohair production and thyroid status of Angora goat kids. *J Agri. Res*, 39: 111-115.
- Yagil R, Z Etzion and J Gannani, 1978. Camel thyroid metabolism: effect of season and dehydration. *J Appl Physiol*, 45: 540-544.
- Zia-ur-Rahman N Ahmad, SA Bukhari, N Akhtar and IU Haq, 2007. Serum hormonal, electrolytes and trace element profiles in the rutting and non-rutting one-humped male camel (*Camelus dromedarius*). *Anim Reprod Sci*, 101: 172-178.